



# Multifaceted link between cancer and inflammation

Gautam SETHI<sup>\*1</sup>, Muthu K. SHANMUGAM<sup>\*</sup>, Lalitha RAMACHANDRAN<sup>\*</sup>, Alan Prem KUMAR<sup>\*†</sup> and Vinay TERGAONKAR<sup>‡1</sup>

<sup>\*</sup>Department of Pharmacology, Yong Loo Lin School of Medicine, National University of Singapore, Singapore 117597, Singapore, <sup>†</sup>Cancer Science Institute of Singapore, National University of Singapore, 28 Medical Drive, Singapore 117456, Singapore, and <sup>‡</sup>Institute of Molecular and Cellular Biology (A\*STAR), 61 Biopolis Drive, Singapore 138673, Singapore

## Synopsis

Increasing evidence from epidemiological, preclinical and clinical studies suggests that dysregulated inflammatory response plays a pivotal role in a multitude of chronic ailments including cancer. The molecular mechanism(s) by which chronic inflammation drives cancer initiation and promotion include increased production of pro-inflammatory mediators, such as cytokines, chemokines, reactive oxygen intermediates, increased expression of oncogenes, COX-2 (cyclo-oxygenase-2), 5-LOX (5-lipoxygenase) and MMPs (matrix metalloproteinases), and pro-inflammatory transcription factors such as NF- $\kappa$ B (nuclear factor  $\kappa$ B), STAT3 (signal transducer and activator of transcription 3), AP-1 (activator protein 1) and HIF-1 $\alpha$  (hypoxia-inducible factor 1 $\alpha$ ) that mediate tumour cell proliferation, transformation, metastasis, survival, invasion, angiogenesis, chemoresistance and radioresistance. These inflammation-associated molecules are activated by a number of environmental and lifestyle-related factors including infectious agents, tobacco, stress, diet, obesity and alcohol, which together are thought to drive as much as 90% of all cancers. The present review will focus primarily on the role of various inflammatory intermediates responsible for tumour initiation and progression, and discuss in detail the critical link between inflammation and cancer.

**Key words:** activator protein 1 (AP-1), hypoxia-inducible factor 1 $\alpha$  (HIF-1 $\alpha$ ), matrix metalloproteinase (MMP), nuclear factor  $\kappa$ B (NF- $\kappa$ B), oncogene, signal transducer and activator of transcription 3 (STAT3)

## INTRODUCTION

Inflammation is a complex process that involves widespread changes in cellular and molecular components of physiology. Although controlled inflammation is a necessary process required for an array of processes including tissue repair, wound healing and for defence against invading foreign pathogens, chronic, uncontrolled inflammation is harmful and has now been linked to a number of human ailments [1,2]. The critical role of chronic inflammation in cancer was first proposed by Rudolf Virchow in 1863, when he observed the presence of leucocytes in neoplastic tissues [3,4]. Virchow postulated that an inflammatory milieu promotes a cellular environment that drives the initiation and development of carcinogenesis [1,5].

Within the tumour microenvironment, a network of various pro-inflammatory mediators participate in a complex signalling process that facilitates extravasations of tumour cells through the stroma, thereby promoting tumour progression [6,7]. While acute inflammation is primarily a self-limiting process and has potential therapeutic consequences, prolonged chronic inflammation is mostly detrimental [2,8]. Chronic inflammation is now dubbed by the popular press as a ‘secret killer’ and has been widely associated with diseases such as atherosclerosis, rheumatoid arthritis, multiple sclerosis, asthma, Alzheimer’s disease and various cancers [1,3,4].

It is a well-accepted paradigm now that environment- and lifestyle-related factors play a critical role in development of 90% of all cancers [4,9]. For example, almost 30% of all cancers have been attributed to tobacco smoke, 35% to diet,

**Abbreviations used:** AP-1, activator protein 1; CCR7, CC chemokine receptor 7; COX-2, cyclo-oxygenase-2; CXCL14, CXC chemokine ligand 14; CXCR, CXC chemokine receptor; ECM, extracellular matrix; EGF, epidermal growth factor; EGFR, EGF receptor; HIF-1 $\alpha$ , hypoxia-inducible factor 1 $\alpha$ ; HPV, human papillomavirus; JAK, Janus kinase; IGF-1, insulin-like growth factor; I $\kappa$ B, inhibitory  $\kappa$ B; IKK, I $\kappa$ B kinase; IL, interleukin; 5-LOX, 5-lipoxygenase; MAPK, mitogen-activated protein kinase; MMP, matrix metalloproteinase; NF- $\kappa$ B, nuclear factor  $\kappa$ B; NSCLC, non-small cell lung cancer; RCC, renal cell carcinoma; ROS, reactive oxygen species; SCC, squamous cell carcinoma; STAT3, signal transducer and activator of transcription 3; TAM, tumour-associated macrophage; TNF, tumour necrosis factor; VEGF, vascular endothelial growth factor.

<sup>1</sup>Correspondence may be addressed to either of these authors (email gautam\_sethi@nuhs.edu.sg or vinayt@imcb.a-star.edu.sg).

14–20% to obesity, 18% to infections and 7% to radiation and environmental pollutants [10]. The molecular mechanism(s) by which these risk factors induce cancer are becoming increasingly evident and one major process that seems to be common between all these risk factors is inflammation. Chronic inflammation acts as a key regulator of tumour promotion and progression by several mechanisms including accelerated cell proliferation, evasion from apoptosis, enhanced angiogenesis and metastasis [11]. The mechanism(s) for cancer development in the presence of chronic inflammation involves the continuous presence of cytokines, chemokines, ROS (reactive oxygen species), oncogenes, COX-2 (cyclo-oxygenase-2), 5-LOX (5-lipoxygenase), MMPs (matrix metalloproteinases) and activation of important transcription factors such as NF- $\kappa$ B (nuclear factor  $\kappa$ B) and STAT3 (signal transducer and activator of transcription 3), AP-1 (activator protein 1) and HIF-1 $\alpha$  (hypoxia-inducible factor 1 $\alpha$ ) [8,12]. In the present review, we will focus on the role of various pro-inflammatory mediators in cancer and provide novel insights into the intricate link between chronic inflammation and cancer.

## ROLE OF TNF (TUMOUR NECROSIS FACTOR) IN INFLAMMATION-DRIVEN CANCERS

TNF $\alpha$  was first isolated as an anticancer cytokine more than two decades ago, but when its antitumour activity was tested on cancer patients, a paradoxical tumour-promoting role of TNF $\alpha$  became apparent [13–15]. At present, the pro-inflammatory role of TNF $\alpha$  has been linked to all steps involved in tumorigenesis, including cellular transformation, survival, proliferation, invasion, angiogenesis and metastasis [15,16].

TNF $\alpha$  has been reported to be produced by a wide variety of tumour cells, including those of B-cell lymphoma [17], megakaryoblastic leukaemia [18], adult T-cell leukaemia [19], breast carcinoma [20], colorectal cancer, lung cancer, SCC (squamous cell carcinoma), pancreatic cancer [21,22], ovarian carcinoma [23], the cervical epithelial cancer [24], glioblastoma [25] and neuroblastoma [26]. The pro-inflammatory potential of TNF $\alpha$  has also been analysed in various animal models of cancer. In a genetic model of liver cancer, TNF $\alpha$  produced by myeloid cells promoted inflammation-associated tumours [27] and also in a chemical-induced model of colorectal cancer, TNF $\alpha$  produced by macrophages has been implicated in inflammation and subsequent tumour development [15]. Endogenous and exogenous TNF $\alpha$  showed an enhancement of metastasis in an experimental fibrosarcoma metastasis model [28]. Elevated levels of TNF $\alpha$  have also been detected in various cancer patients. For example, the TNF $\alpha$  gene was found to be expressed in 45 of 63 biopsies of human epithelial ovarian cancer [23]. Moreover, it has been found that, in CLL (chronic lymphocytic leukaemia) patients, TNF $\alpha$  level was significantly higher as compared with the healthy con-

trol population and it also acted as a predictor of patient survival [29]. Thus, novel strategies that neutralize systemic TNF $\alpha$  may be useful in cancer treatment and prevention.

## ROLE OF IL (INTERLEUKIN) IN INFLAMMATION AND CANCER

Several ILs have been linked with inflammation and subsequent cancer development. These ILs include IL-1, IL-6, IL-8 and IL-17. IL-1 $\alpha$ , which is expressed in both normal tissue and several tumour cells, is a regulatory cytokine that can induce the activation of transcription factors, including NF- $\kappa$ B and AP-1, and promote the expression of various genes involved in cell survival, proliferation and angiogenesis [30]. Also, direct evidence for the role of IL-1 $\beta$  in human cancer has been found in multiple myeloma. IL-1 $\beta$  when released by myeloma cells can induce the production of IL-6 by bone marrow stromal cells and function as an autocrine growth factor for myeloma cells [31]. IL-1 $\beta$  also up-regulates HIF-1 $\alpha$  protein through a classical inflammatory signalling pathway involving NF- $\kappa$ B and COX-2, culminating in up-regulation of VEGF (vascular endothelial growth factor), a potent angiogenic factor required for tumour growth and metastasis [32]. In another study, surgical removal of the ovarian tumour and resolution of ascites in patient was found to be directly associated with decrease in serum levels of IL-1 $\beta$  [33].

IL-6 is another major pro-inflammatory cytokine that has been implicated in inflammation-associated carcinogenesis [34,35]. IL-6 modulates the expression of genes involved in proliferation, survival and angiogenesis via the JAK (Janus kinase)–STAT signalling pathway [36]. RCC (renal cell carcinoma) cell lines containing mutant p53 have been found to produce higher levels of IL-6 than those containing wild-type p53 [37]. Moreover, the analysis of biopsy specimens from inflammation-associated gastric cancers has revealed that the levels of IL-1 $\beta$  and IL-6 are highly elevated in tumours as compared with adjacent normal mucosa [38]. An overproduction of IL-6, indicated by increased plasma CRP (C-reactive protein) levels, has also been found in 37% of multiple myeloma patients at diagnosis and is associated with disease aggressiveness, myeloma-cell proliferation and poor prognosis [39]. Increased serum levels of IL-6 have been observed to be positively correlated with tumour burden in colorectal cancer patients with high significance [40]. In another study, inflammatory markers were measured at baseline in 52 patients with stage IV colorectal cancer, and significantly elevated levels of IL-6 and gp130 were observed in these patients and inflammatory markers paralleled clinical outcome [41].

Constitutive expression of IL-8 mRNA and secreted IL-8 protein has been observed in various tumour cell lines and animal models, thus suggesting that IL-8 secretion could be a key factor involved in proliferation, angiogenesis and metastasis of cancer cells [42]. It has been reported that acidic pH can induce elevation in IL-8 expression in human ovarian cancer cells and transcription factors; AP-1 and NF- $\kappa$ B were found to be responsible for this

process [43]. Huang et al. [44] have further found that the neutralizing antibodies to IL-8 can inhibit angiogenesis, tumour growth and metastasis of human melanoma, suggesting the potential utility of anti-IL-8 as a modality to treat melanoma and other solid tumours either alone or in combination with conventional chemotherapy or other antitumour agents. In another report, tumour-derived IL-8 has been shown to induce the differentiation and activation of osteoclasts, underpinning the characteristic osteolytic metastasis of breast cancer cells that have disseminated to the bone [45]. Furthermore, Maxwell et al. [46] determined whether hypoxia can increase IL-8 and IL-8 receptor expression in prostate cancer cells and whether this contributes to a survival advantage in hypoxic cells. Indeed, they found that IL-8, CXCR1 (CXC chemokine receptor 1) and CXCR2 mRNA expression in prostate cancer PC3 cells was up-regulated in response to hypoxia in a time-dependent manner. They also found that the inhibition of IL-8 signalling potentiated etoposide-induced cell death in hypoxic PC3 cells [46]. These results indicate that IL-8 signalling confers a survival advantage to hypoxic prostate cancer cells, and therefore strategies to inhibit IL-8 signalling may sensitize hypoxic tumour cells to conventional treatments. IL-17, another important cytokine, has also been found to act as a growth factor in cutaneous T-cell lymphoma and a key regulator of angiogenesis [47]. IL-17-overexpressing human cervical cancer [48], fibrosarcoma [49] and human NSCLC (non-small cell lung cancer) preferentially exhibit higher oncogenic growth *in vivo* [50].

## ROLE OF CHEMOKINES IN INFLAMMATION AND CANCER

Chemokines are soluble chemotactic cytokines that are grouped into four classes based on the positions of key cysteine residues: C, CC, CXC and CX3C [8,51,52]. Several studies have reported the involvement of chemokines and chemokine receptors in cell proliferation, migration, and invasion and metastasis of different types of tumours [53–55].

The chemokine receptors CXCR4 and CCR7 (CC chemokine receptor 7) are highly expressed in human breast cancer cells, malignant breast tumours and metastasis [56]. In breast cancer cells, signalling through CXCR4 or CCR7 mediates actin polymerization and pseudopodia formation and subsequently induces chemotactic and invasive responses [56]. It has been reported that CXCR4 and SDF-1 (stromal-cell-derived factor 1) induces proliferation in ovarian cancer cells, and this correlated with EGFR [EGF (epidermal growth factor) receptor] transactivation. The functional chemokine receptor CCR3 has been shown to be up-regulated in human RCC [57]. CXCL14 (CXC chemokine ligand 14) [BRAK (breast and kidney chemokine)] RNA expression has been observed in normal and tumour prostate epithelium and focally in stromal cells adjacent to cancer [58]. *In vivo*, neutralizing the interactions of CXCL12/CXCR4 significantly impairs metastasis of breast cancer cells to regional lymph nodes and lung [59]. Thus chemokines and their receptors have a critical role in

determining the metastatic destination of tumour cells. A list of various ILs and chemokines associated with cancer initiation and promotion is briefly summarized in Table 1.

## ROLE OF ONCOGENES IN INFLAMMATION-DRIVEN CANCER

Oncogenes are altered versions of normal cellular genes, the so-called proto-oncogenes, involved in the regulation of cell growth [60,61]. Recently, it has become increasingly evident that pleiotropic effects of oncogenes also include the induction of a pro-tumour microenvironment, through the persistent promotion of an inflammatory milieu [61–63]. For example, Liu et al. [64] have shown that HRAS- and KRAS-G12V induce the expression of various cytokines, including IL-1 $\alpha$ , IL-1 $\beta$ , IL-6, CXCL8 and IL-11 in human ovarian cells [64]. Moreover, transcription factor NF- $\kappa$ B is activated in Ras-transformed ovarian epithelial cells and this activation is responsible for the increased expression of CXCL8 [65]. Furthermore, Ancrile et al. [66] have demonstrated that IL-6 acts downstream of Ras in a paracrine fashion to promote angiogenesis. Recent reports indicate that Myc oncogene can also orchestrate a complex inflammatory program [67]. Myc activation in  $\beta$ -cells rapidly induces the expression and release of the pro-inflammatory cytokine IL-1 $\beta$  that, in turn, mediates the release of VEGF-A from mast cells and onset of tumour angiogenesis [68]. Mast cell activation is required not only for angiogenesis during outgrowth of Myc-dependent islet tumours but also for tumour maintenance and the inhibitors of mast cell function trigger hypoxia and cell death of tumour and endothelial cells [69]. Moreover, four mutually exclusive genetic lesions have been identified in papillary thyroid carcinoma, covering approx. 80% of the cases: rearrangements of *Ret* or *Trk* genes and activating mutation of *Ras* or *Braf* genes [70]. Thus, several oncogene-driven inflammatory pathways are activated in various human cancers and are likely to play a key role in various stages of carcinogenesis.

## ROLE OF OXIDATIVE STRESS IN CHRONIC INFLAMMATION AND CANCER

Reactive oxygen intermediates, also generically referred to as oxidants, are derivatives of molecular oxygen such as superoxide, H<sub>2</sub>O<sub>2</sub>, hypochlorous acid, singlet oxygen and the hydroxyl radical [71–73]. Chronic inflammation is often accompanied by increased production of tissue reactive oxygen intermediates [74]. ROS can alter signal transduction cascades as well as induce changes in transcription factors such as NF- $\kappa$ B, NF-E2/rf2 or Nrf2 (nuclear factor erythroid 2/related factor 2) and AP-1 that mediate immediate cellular stress responses [75,76]. The

**Table 1 Role of ILs and chemokines in cancer**MIP-3 $\alpha$ , macrophage inflammatory protein-3 $\alpha$ .

Cancer type	Inflammatory mediator	Mechanism(s)	Reference
Bladder cancer	IL-6	Transformation	[193]
Multiple myeloma	IL-6 poly	Proliferation	[194]
Colorectal cancer	IL-6	Increased risk	[195]
Melanoma	IL-18	Metastasis	[196]
Pancreatic carcinoma	IL-1 $\alpha$	Metastasis	[197]
Prostate cancer	IL-8 poly	Angiogenesis	[198]
Lung carcinoma	IL-1 $\alpha$	Angiogenesis	[32]
Melanoma	IL-8	Tumour growth	[199]
Glioblastoma	IL-8	Angiogenesis	[200]
RCC	IL-6	Autocrine growth	[37]
Pancreatic carcinoma	IL-1 $\beta$	Chemoresistance	[201]
Ovarian tumours	IL-8	Disease progression	[202]
Tumour	IL-8	Growth, angiogenesis	[203]
Lung carcinoma	IL-1 $\beta$	Growth	[204]
Breast cancer	CXCR4 and CCR7	Metastasis	[56]
Melanoma	CXCR4 and CCR7,	Metastasis	[56]
Ovarian carcinoma	CXCR4/CXCL12	Invasion and growth	[43]
RCC	CCR3	Higher risk	[57]
Pancreatic carcinoma	MIP-3 $\alpha$ and CCR6	Cell invasion	[205]
Ovarian carcinoma	CXCR4 and SDF1	Proliferation	[206]
Prostate carcinoma	CXCL14	Inhibits tumour growth	[58]

pro-neoplastic activity of ROS is mainly due to their ability to cause DNA damage [77]. Oxidative damage to DNA has also been linked to aflatoxin B-induced p53 and Ras gene mutations in hepatocarcinogenesis [78] and in UV-induced mouse and human skin cancers [79]. Agents that either scavenge reactive oxygen intermediates or prevent their formation inhibit the induction of DNA damage, mutagenesis and transformation by inflammatory phagocytes. This forms the basis for the theory that dietary antioxidants can inhibit the development or progression of cancer [80–82].

## OVEREXPRESSION OF COX CAN MEDIATE INFLAMMATION-ASSOCIATED CANCERS

COX-2, an inducible enzyme regulated by NF- $\kappa$ B, is known to mediate tumorigenesis [11,83]. COX-2, the inducible isoform of prostaglandin H synthase, has been implicated in the growth and progression of a variety of human cancers. COX-2 has been shown to regulate colorectal cancer-induced angiogenesis by two mechanisms: COX-2 can modulate the production of angiogenic factors by colon cancer cells, while COX-1 regulates angiogenesis in endothelial cells. It has been found that COX-2 and mPGES (membrane-associated prostaglandin E synthase) were induced in the COX-1-expressing fibroblasts in human familial

adenomatous polyposis polyps [84]. Administration of the COX-2-selective inhibitor rofecoxib or the functional inactivation of the COX-2 in adenomatous polyposis coli knockout mice, a murine model of human adenomatous polyposis, reduced the number and the size of intestinal polyps [85,86], thereby indicating the correlation between the abnormal up-regulation of COX-2 and tumorigenesis.

COX-2 expression in human tumours can be induced by growth factors, cytokines, oncogenes and other factors. For example, IL-1 $\beta$  has been reported to up-regulate COX-2 in human colorectal cancer cells via multiple signalling pathways [87]. COX-2 has also been implicated in the progression of human lung adenocarcinoma. Steady-state levels of COX-2 mRNA were high in well-differentiated adenocarcinoma samples but low in poorly differentiated adenocarcinoma, SCC and small cell lung cancer. COX-2 overexpression enhanced the *in vitro* expression of both CXC ligand CXCL8 and CXCL5, NSCLC angiogenic peptides in the NSCLC cell lines [88]. COX-2 expression was observed to be strong in the SCCs and weak in oesophageal ADCs (adenocarcinomas) [89]. COX-2 expression levels in tumour specimens from patients with low- and high-grade astrocytomas indicated a correlation between the percentage of COX-2 expression and patient survival [90]. Overexpression of COX-2 is also associated with a poor prognosis in patients with SCC of the uterine cervix treated with radiation and concurrent chemotherapy [91]. Levels of COX-2 expression were also found to be a significant prognostic factor for patients with multiple myeloma [92]. Overall survival of those patients

with negative or weak-to-moderate COX-2 expression was significantly better than that of patients with strong COX-2 immunoreactivity. These findings indicate that high COX-2 expression in tumour cells is associated with clinically more aggressive tumours and is a strong predictor of poor survival.

OVEREXPRESSION OF 5-LOX LINKS INFLAMMATION AND CANCER

5-LOX is a key enzyme in the metabolism of arachidonic acid to leukotrienes [93]. Several studies suggest that there is a link between 5-LOX and carcinogenesis in humans and animals [93–95]. Abundance of the mRNA for arachidonate 5-LOX, which is the rate-limiting enzyme in leukotriene synthesis, has been investigated in a series of human brain tumours [96]. The 5-LOX transcript is expressed in human brain tumours and the 5-LOX gene product may play a role in human tumour-induced brain oedema [96].

Studies also indicate that the exposure to the mainstream smoke of unfiltered cigarettes enhanced the 5-LOX protein expression in the inflammation-associated colonic adenomas [97]. Such expression was accompanied by an up-regulation of MMP-2 and VEGF, the key angiogenic factors for tumorigenesis and 5-LOX inhibitors were found to decrease the incidence of colonic adenoma formation and reduced angiogenesis, MMP-2 activity and VEGF protein expression [97]. In addition, the increased expression of 5-LOX has been linked with the progression and development of cancer of the pancreas [98], breast [99] and kidney [100].

ROLE OF MMP IN INFLAMMATION AND CANCER

MMPs are a multigene family of zinc-dependent endopeptidases that share a similar structure and which collectively have the capacity to degrade ECM (extracellular matrix) [101]. MMPs are now also implicated in the EMT (epithelial to mesenchymal transition), a hallmark of cancer progression to metastasis [102]. It has been observed that MMP-9 is a potent regulator of the angiogenic switch in a pancreatic tumour model [103]. MMP-9 is up-regulated in angiogenic dysplasias and invasive cancers of the epidermis in a mouse model of multi-stage tumorigenesis elicited by HPV16 (human papillomavirus 16) oncogenes [104]. In gene expression profiles associated with poor outcome of patients with breast tumours, two of the 70 genes identified were found to be MMP-1 and MMP-9 [105]. In another study, patient survival, gene overexpression and RNAi (RNA interference) validation data showed that MMP-1 is the second most important gene in a 95-gene expression profile in determining the metastatic potential of breast cancer to produce lung metastases [106]. Expression of MMP-9 has also been correlated with prognosis, aggressiveness

Table 2 Role of inflammatory enzymes (COX2, 5-LOX and MMP-9) in cancer  
AC, adenocarcinoma.

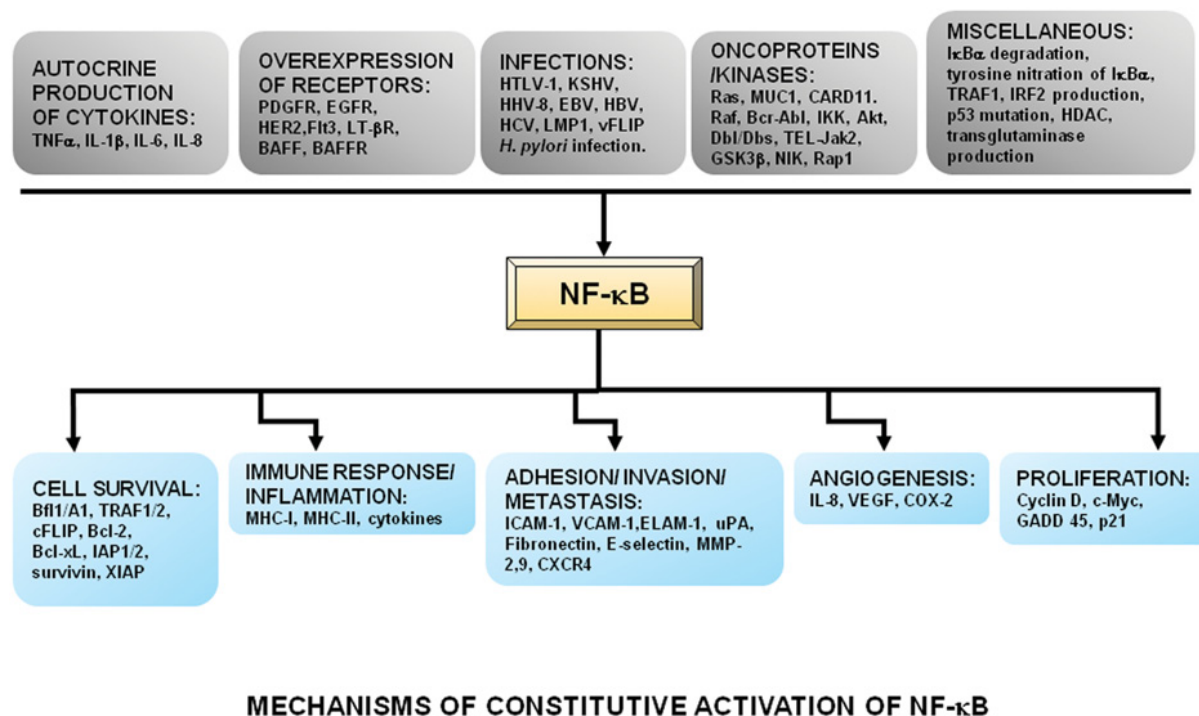
Tumour	Enzyme	References
Breast cancer	COX-2	[207,208]
Glioma	COX-2	[90]
Prostate cancer	COX-2	[209]
Melanoma	COX-2	[210,211]
Oesophageal adenocarcinoma	COX-2	[212]
Oesophageal SCC and AC	COX-2	[89]
Urinary bladder	COX-2	[213]
Pancreatic cancer	COX-2	[214]
Head and neck SCC	COX-2	[215]
Lung carcinoma	COX-2	[216,217]
Gastric carcinoma	COX-2	[218]
Colorectal cancer	COX-2	[84,219]
Brain tumours	5-LOX	[96,220]
Colorectal cancer	COX-2 and 5-LOX	[221]
Pancreatic cancer	MMP-9	[103]
Skin cancer	MMP-9	[104]

and survival in cancer of the lung [107], stomach [108] and oesophagus [109], RCC and in NHL (non-Hodgkin’s lymphoma) [110]. A role of COX-2, 5-LOX, and MMPs in cancer is briefly summarized in Table 2.

ROLE OF TRANSCRIPTION FACTOR NF-κB IN CHRONIC INFLAMMATION AND CANCER

The transcription factor NF-κB, first discovered by David Baltimore in 1986, is present in the nucleus and binds the promoter of immunoglobulin κ chain in B-cells. In mammalian cells, the NF-κB family of transcription factors is composed of homodimers and heterodimers derived from five distinct subunits, RelA (p65), c-Rel, RelB, p50 (NF-κB1) and p52 (NF-κB2). All family members share a highly conserved RHD (Rel homology domain; ~300 amino acids) responsible for DNA binding, dimerization domain and interaction with IκBs (inhibitory κBs), the intracellular inhibitor of NF-κB [111–113]. In unstimulated cells, the majority of NF-κB complexes are predominantly cytoplasmic and in an inactive form due to their binding to the IκB family of proteins that prevent DNA binding and as a consequence prevent nuclear accumulation [114]. Generally, the inactive NF-κB/IκBα complex is activated by phosphorylation on two conserved serine residues within the N-terminal domain of the IκB proteins. Phosphorylation of these conserved serine residues in response to stimulation leads to the immediate polyubiquitination of IκB proteins by the SCF-β-TrCP (transducin repeat-containing protein-β-transducin repeat-containing protein) complex. This modification subsequently targets IκB proteins for rapid degradation by the 26S proteasome [115]. Activation of





**Figure 1** Mechanisms of constitutive activation of NF- $\kappa$ B

Abbreviations: BAFF, B-cell activating factor belonging to the TNF family; BAFFR, B-cell activating factor belonging to the TNF family receptor; CARD11, caspase recruitment domain family 11; Dbp/Dbs, transforming protein isolated from diffuse B-cell lymphoma; EBV, Epstein Bar virus; ELAM-1, endothelial cell leucocyte adhesion molecule 1; FcγR, fms-related tyrosine kinase 3; GADD, growth arrest and DNA-damage inducible; GSK3 $\beta$ , glycogen synthase kinase 3 $\beta$ ; HBV, hepatitis B virus; HCV, hepatitis C virus; HDAC, histone deacetylase; HER2, erythroblastic leukaemia viral oncogene; HHV-8, human herpes virus 8; HTLV-1, human T-cell leukaemia virus type 1; ICAM-1, intracellular adhesion molecule 1; IRF2, interferon regulatory factor 2; KSHV, Kaposi's sarcoma-associated herpes virus; LMP1, latent membrane protein 1; LT- $\beta$ R, lymphotxin  $\beta$  receptor; MUC1, mucin 1; PDGFR, platelet-derived growth factor receptor; TEL-Jak2, telomere maintenance-Janus kinase 2; TRAF, TNF-receptor-associated factor; uPA, urokinase plasminogen activator; VCAM-1, vascular cell adhesion molecule 1; vFLIP, viral FADD-like interleukin-1 $\beta$ -converting enzyme (FLICE)/caspase-8-inhibitory protein; XIAP, X-linked inhibitor of apoptosis.

the NF- $\kappa$ B signalling cascade is a consequence of degradation of I $\kappa$ B proteins, allowing nuclear accumulation of NF- $\kappa$ B, due to DNA binding [116–119]. NF- $\kappa$ B is activated by many divergent stimuli, including pro-inflammatory cytokines (e.g. TNF $\alpha$ , IL-1), T- and B-cell mitogens, bacteria, LPS (lipopolysaccharide), viruses, viral proteins, double-stranded RNA and physical and chemical stresses. Activated NF- $\kappa$ B binds to specific DNA sequences in target genes, designated as  $\kappa$ B elements, and regulates transcription of over 400 genes involved in inflammation, immunoregulation, tumour cell proliferation, invasion, metastasis, angiogenesis, chemoresistance and radioresistance [120–124].

Numerous studies have indicated that tumour cells exhibit constitutive production of the pro-inflammatory cytokines TNF $\alpha$ , IL-1 $\alpha$ , IL-6, GM-CSF (granulocyte/macrophage colony-stimulating factor) and KC (keratinocyte chemoattractant) [2,125]. Production of tumour-promoting cytokines by immune/inflammatory cells that activate NF- $\kappa$ B, along with other transcription factors such as AP-1 and STAT3 in premalignant cells to induce genes that stimulate cell proliferation and survival, is a major tumour-promoting mechanism [2,125]. For instance, inhibition of TNF $\alpha$

production by non-parenchymal cells (Kupffer and endothelial cells) prevented NF- $\kappa$ B activation in hepatocytes and in early tumours and reduced tumour multiplicity [27]. Greten et al. [126] reported that deleting IKK $\beta$  (I $\kappa$ B kinase  $\beta$ ) in myeloid cells caused suppression of NF- $\kappa$ B, activation and diminished expression of inflammatory cytokines, thus leading to a significant decrease in tumour size. The host environment promotes the constitutive activation of NF- $\kappa$ B and pro-inflammatory cytokine expression during metastatic tumour progression of various cancers [113,127,128].

What causes the constitutive activation of NF- $\kappa$ B in various tumour cells is not fully understood. Many different mechanism(s) have been described, including overexpression of growth factor receptors, mutation of I $\kappa$ B $\alpha$  such that it cannot bind to NF- $\kappa$ B, constitutive activation of Ras protein, high proteolytic activity directed to I $\kappa$ B $\alpha$ , and autocrine secretion of inflammatory cytokines (Figure 1). It has also been shown that I $\kappa$ B proteins do not bind and export NF- $\kappa$ B that is phosphorylated at p65. Indeed phosphatases of p65 such as WIP1 (wild-type p53-induced phosphatase) have recently been identified with removed phosphates

from p65 and make NF- $\kappa$ B more submissive to I $\kappa$ B-mediated nuclear export [129]. Constitutive activation of NF- $\kappa$ B also has been linked to chemoresistance and radioresistance in various tumour cell lines and in animal models [113,124]. It is also well known that it blocks the function of p53 tumour suppressor by causing its degradation [130–132]. Activation of IKKs in response to inflammatory stimuli has also been shown to deregulate cell cycle [133]. Thus the activation of NF- $\kappa$ B represents the central event in linking the process of chronic inflammation to different aspects of tumorigenesis. Indeed agents that simultaneously target the p53 and NF- $\kappa$ B pathway should be developed further in the treatment of cancers [134–136].

An association between the development of cancer and inflammation is further strengthened by studies of the role of NF- $\kappa$ B in tumour-infiltrating leucocytes [137]. For example, myeloid-lineage-specific inactivation of the gene encoding IKK $\beta$  was found to inhibit cancer-related inflammation in the intestine, as well as colitis-associated cancer, providing evidence that inflammatory cells are involved in carcinogenesis [126]. Defective NF- $\kappa$ B has also been reported in T-lymphocytes of patients with RCC [138]. In established advanced tumours, which typically have an inflammatory milieu [139], TAMs (tumour-associated macrophages) have delayed and defective NF- $\kappa$ B activation [140]. Inhibition of NF- $\kappa$ B activation in TAMs has also been reported to correlate with impaired expression of NF- $\kappa$ B-dependent inflammatory functions [141] and to exhibit the alternatively activated, ‘M2’, phenotype [137]. Evidence suggests that homodimers of the p50 subunit of NF- $\kappa$ B (a negative regulator of the NF- $\kappa$ B pathway) are responsible for this slow activation of NF- $\kappa$ B in TAMs and for the pro-tumour phenotype of these cells [142]. Thus, NF- $\kappa$ B seems to function as a ‘rheostat’ whose function can be tuned to different levels, predisposing individuals towards developing cancer, and enables TAMs to maintain the inflammatory milieu [137]. Although several experimental and clinical results clearly indicate inflammation having a pro-tumour consequence, some reports also demonstrate the inverse. For example, a marked chronic inflammatory response is not associated with an increased risk of developing melanoma [143]. Also, in certain tumours, the presence of inflammatory cells is associated with better prognosis [144]. These observations appear to reveal that inflammatory cells can destroy tumour cells, in addition to normal tissue cells. Taken together, evidence indicates that NF- $\kappa$ B is an important determinant of the balance between the pro-tumour and anti-tumour properties of macrophages [142,145] and thus NF- $\kappa$ B could be targeted to ‘re-educate’ tumour-promoting macrophages towards an anti-tumour role [145].

## ROLE OF STAT3 IN INFLAMMATION AND CANCER

STAT3 was originally identified as a DNA-binding protein that responds to stimulation by EGF and IL-6 and has an important role in their signalling [146,147]. On activation, STAT3 undergoes

phosphorylation-induced homodimerization, leading to nuclear translocation, DNA binding and subsequent gene transcription [148]. The phosphorylation is mediated through the activation of non-receptor protein tyrosine kinases called JAKs. JAK1, JAK2, JAK3 and TYK2 have been implicated in the activation of STAT3 [147,149]. Constitutive activation of STAT3 has been observed in many kinds of solid tumours and haematological malignancies [4,150] and this persistently active STAT3 is thought to contribute to oncogenesis by modulating the expression of a variety of genes involved in cell proliferation, invasion, metastasis and angiogenesis [151,152].

Chronic inflammatory conditions that drive carcinogenesis can also be attributed to genetic alterations that directly affect the STAT3 pathway [149]. The importance of constitutively active mutations in GP130, which encodes a subunit of the IL-6 receptor, has been demonstrated in human inflammatory HCC (haemofiltrate CC chemokine) [153]. A critical role for STAT3 in inflammation-induced adenocarcinomas was also demonstrated using a transgenic mouse model with a constitutively active GP130 in epithelial cells [154]. Studies in mice with GP130 mutations demonstrated that an increase in GP130 and STAT3 signalling led to inflammation-associated gastric tumorigenesis [155]. Several infectious agents also exert their tumorigenic effects through STAT3 activation and depend on STAT3 for their oncogenic potential [149]. For instance, infection with *Helicobacter pylori*, which is associated with gastric cancer, activates STAT3 through its cytotoxin-associated gene A in host cells [156]. In addition, a critical role of STAT3 activation in mediating UV-light-induced skin cancer in a transgenic mouse model and cigarette-smoke-associated cancer development has also been demonstrated [157,158].

STAT3 can also act in close liaison with NF- $\kappa$ B to mediate various steps involved in initiation, promotion and development of cancer [159]. Moreover, NF- $\kappa$ B and STAT3 control both distinct and overlapping groups of genes involved during tumorigenesis [149]. Global profiling of STAT3-dependent genes in mouse lung cells revealed a large number of genes whose expression is controlled by STAT3, among which a number of typical NF- $\kappa$ B target genes are also present [160]. Furthermore, in a recent study, it was demonstrated that obesity-promoted hepatocellular carcinoma development was dependent on enhanced production of the tumour-promoting cytokines IL-6 and TNF $\alpha$ , which cause hepatic inflammation and activation of the STAT3 [161]. Thus STAT3 activation pathway also is an important contributor to inflammation-induced cancers, making it an attractive target for treating and/or preventing inflammation.

## ROLE OF AP-1 IN INFLAMMATION AND CANCER

The transcription factor AP-1 produced by 18 different dimeric combinations of proteins from the Jun (c-Jun, JunB and JunD) and Fos (c-Fos, FosB, Fra-1 and Fra-2) families, plays a critical role

in variety of cellular processes, including inflammation, proliferation, differentiation and apoptosis [162–164]. When activated, AP-1 recognizes and binds to the TRE (TPA response element) or cAMP response element within the promoter region of target genes [165]. Activation usually occurs both transcriptionally and post-translationally in response to a broad range of external stimuli, including growth factors, pro-inflammatory cytokines, chemokines, ECM and is mediated predominantly through the MAPK (mitogen-activated protein kinase) [ERK (extracellular-signal-regulated kinase), JNK (c-Jun N-terminal kinase) and p38 MAPK] cascade [166,167]. In addition to being activated by oncogenic signal transduction cascades, AP-1 is itself strongly oncogenic [168]. Endogenous *c-fos* and *c-jun* are also oncogenes as indicated by their potential to morphologically transform murine fibroblasts, causing density- and anchorage-independent growth in these cells [169,170]. Moreover, inhibition of Fos and Jun expression in murine fibroblasts and erythroleukaemia cells has indicated that AP-1 is required for cell proliferation and cell-cycle progression [164].

AP-1 is also overexpressed in a large number of tumours and transformed cell lines and targeted inhibition of its activity in these model systems suggest a pivotal role for AP-1 in oncogenic transformation and progression [163,164]. Dominant-negative constructs of *c-fos* and *c-jun* can reverse the transformed phenotype induced by activated Ras and also inhibit the invasiveness and tumorigenesis of keratinocytes [171]. Several AP-1 target genes are also implicated in the invasive phenotype, including the MMPs MMP-1, MMP-3 and MMP-9 [172], the ECM-associated protein osteonectin/SPARC (secreted protein acidic and rich in cysteine) [173], the PKC (protein kinase C) substrate SSeCKS (Src-suppressed C-kinase substrate) [174] and the angiogenic factor, autotaxin [175]. Interestingly, suppression of *c-jun* activity by using a dominant-negative *c-jun* in basal keratinocytes or conditional inactivation of *c-jun* in the liver resulted in the inhibition of the development of chemically induced papillomas and liver tumours respectively [176,177]. Moreover, using mice overexpressing *c-fos*, Wang et al. [178] showed an intimate relationship between *c-fos* expression levels and chondrogenic tumour development. Furthermore, AP-1 has also been found to interact with pro-inflammatory transcription factor NF- $\kappa$ B, and the dominant-negative Jun has been reported to inhibit both AP-1 and NF- $\kappa$ B activity in HPV-immortalized human keratinocytes [179]. Thus AP-1 acts as a master regulator of gene expression in response to oncogenic signal transduction cascades in a wide variety of tumour cell and animal models and can be considered as an important target for novel anti-cancer therapies.

## ROLE OF HIF-1 $\alpha$ IN INFLAMMATION AND CANCER

HIF-1 is a heterodimeric transcriptional complex composed of an  $\alpha$ -subunit and a  $\beta$ -subunit [180,181]. The HIF-1 $\alpha$  subunit

is generally unstable and undergoes proteasomal degradation in normoxia, whereas the  $\beta$ -subunit is permanently present in nuclei irrespective of the state of oxygenation [182]. Recent studies have shown that a number of peptidic and non-peptidic mediators of inflammation can activate HIF-1 $\alpha$  even under normoxic conditions [183]. These include cytokines, hormones such as insulin or IGF-1 (insulin-like growth factor 1) and IGF-2, and vasoactive peptides, such as angiotensin II [183]. Among various cytokines, TNF $\alpha$  and IL-1 $\beta$  were first shown to increase HIF-1 $\alpha$  activity in the human hepatoma cell line HepG2 [184]. HIF-1 $\alpha$  stimulates the expression of several genes encoding the proteins that promote inflammatory reactions. These include erythropoietin, VEGF and VEGF receptor, iNOS (inducible nitric oxide synthase), COX-2, glucose transporters and a number of glycolytic enzymes [185,186]. Moreover, the accumulation of HIF-1 $\alpha$  in the absence of apparent hypoxic stimulation has been demonstrated in a number of different cancers, in contrast with benign tumours and normal tissues. For example, immunohistochemical analyses of tissue sections have shown HIF-1 $\alpha$  to be highly expressed in many tumour types including pancreatic, head and neck, breast, renal, ovarian, bladder, brain, colorectal and prostate [185]. HIF-1 $\alpha$  overexpression has also been found to correlate with increased angiogenesis and metastasis and thus can be used as a marker to predict outcome in patients with metastatic cancers [181]. Thus, targeting the HIF-1 $\alpha$  pathway provides an attractive strategy to treat various hypoxic and metastatic tumours.

## CONCLUSION AND PERSPECTIVES

There is growing evidence, as described above, which is highly suggestive that chronic inflammation is a critical mediator of various aspects of development of cancer. It is becoming increasingly clear that chronic inflammation contributes to carcinogenesis at all three stages: initiation, proliferation and progression. Some of the agents that have the potential to suppress these pro-inflammatory mediators and are being tested include TNF $\alpha$  blockers (such as thalidomide, enbrel, humira and remicade), IL-1 blockers (canakinumab and anakinra), NF- $\kappa$ B inhibitors (such as curcumin, resveratrol and roscovitine) and COX-2 inhibitors (such as celecoxib). However, while most evidence discussed above indicates that pro-inflammatory cytokines, enzymes, oncogenes and transcription factors play a pivotal role in mediating tumorigenesis, the existing literature also suggests that inhibition of pro-inflammatory pathways is not always beneficial. For example, in a skin cancer mouse model, the pro-inflammatory transcription factor NF- $\kappa$ B has been reported to inhibit tumour formation [187]. Furthermore, in Mdr2-knockout mice, bile duct tumours are rarely found, despite extensive inflammation, NF- $\kappa$ B activation and abundant proliferation of bile ducts in portal spaces [27]. Another recent report indicates that inhibition of NF- $\kappa$ B activation can accelerate hepatocellular carcinoma development and enhance proliferation of tumour-initiating cells [188]. And finally, administration of TNF $\alpha$  blockers to patients



with rheumatoid arthritis have been found to increase the risk for developing lymphomas [189], thereby suggesting that inhibition of pro-inflammatory pathways can act as a double-edge sword.

Therefore novel strategies such as identification of specific adaptors of IKK complex like ELKS [190] and Rap1 [191] will allow development of better inhibitors of IKK and hence NF- $\kappa$ B which are less likely to have adverse side effects. Moreover, genetic studies in patients with hyper-IgE syndrome identified dominant-negative STAT3 gene mutations as the probable cause of the disease in few patients [192]. Thus a detailed elucidation of the underlying mechanism(s) will help us to better understand the interaction between tumour cells and their inflammatory microenvironment, and consequently how to interfere and block such pro-tumour biomarkers with minimum toxic effects. Targeted therapies that can interfere with the recruitment of bone-marrow-derived cells or specifically directed at specific components of the tumour microenvironment can also be utilized in the future as treatment regimens for inflammation-driven cancers.

#### FUNDING

Our own work was supported by the NUS Academic Research Fund [grant number R-184-000-177-112 and R-184-000-170-112 (to G.S.)], National Kidney Foundation [grant number R-184-000-196-592 (to G.S.)], the National Medical Research Council of Singapore [grant number R-713-000-124-213 (to A.P.K.)] and Cancer Science Institute of Singapore, Experimental Therapeutics I Program [grant number R-713-001-011-271 (to A.P.K.)].

#### REFERENCES

- Mantovani, A. (2009) Cancer: inflaming metastasis. *Nature* 457, 36–37
- Grivennikov, S. I., Greten, F. R. and Karin, M. (2010) Immunity, inflammation, and cancer. *Cell* 140, 883–899
- Mantovani, A., Allavena, P., Sica, A. and Balkwill, F. (2008) Cancer-related inflammation. *Nature* 454, 436–444
- Aggarwal, B. B., Vijayalekshmi, R. V. and Sung, B. (2009) Targeting inflammatory pathways for prevention and therapy of cancer: short-term friend, long-term foe. *Clin. Cancer Res.* 15, 425–430
- Demaria, S., Pikarsky, E., Karin, M., Coussens, L. M., Chen, Y. C., El-Omar, E. M., Trinchieri, G., Dubinett, S. M., Mao, J. T., Szabo, E. et al. (2010) Cancer and inflammation: promise for biologic therapy. *J. Immunother.* 33, 335–351
- Colotta, F., Allavena, P., Sica, A., Garlanda, C. and Mantovani, A. (2009) Cancer-related inflammation, the seventh hallmark of cancer: links to genetic instability. *Carcinogenesis* 30, 1073–1081
- Balkwill, F. and Mantovani, A. (2001) Inflammation and cancer: back to Virchow? *Lancet* 357, 539–545
- Aggarwal, B. B. and Gehlot, P. (2009) Inflammation and cancer: how friendly is the relationship for cancer patients? *Curr. Opin. Pharmacol.* 9, 351–369
- Karin, M., Lawrence, T. and Nizet, V. (2006) Innate immunity gone awry: linking microbial infections to chronic inflammation and cancer. *Cell* 124, 823–835
- Anand, P., Kunnumakkara, A. B., Sundaram, C., Harikumar, K. B., Tharakan, S. T., Lai, O. S., Sung, B. and Aggarwal, B. B. (2008) Cancer is a preventable disease that requires major lifestyle changes. *Pharm. Res.* 25, 2097–2116
- Aggarwal, B. B., Shishodia, S., Sandur, S. K., Pandey, M. K. and Sethi, G. (2006) Inflammation and cancer: how hot is the link? *Biochem. Pharmacol.* 72, 1605–1621
- Balkwill, F. and Mantovani, A. (2010) Cancer and inflammation: implications for pharmacology and therapeutics. *Clin. Pharmacol. Ther.* 87, 401–406
- Aggarwal, B. (2003) Signalling pathways of the TNF superfamily: a double-edged sword. *Nat. Rev. Immunol.* 3, 745–756
- Sethi, G., Sung, B. and Aggarwal, B. B. (2008) TNF: a master switch for inflammation to cancer. *Front. Biosci.* 13, 5094–5107
- Balkwill, F. (2009) Tumour necrosis factor and cancer. *Nat. Rev. Cancer* 9, 361–371
- Balkwill, F. (2006) TNF- $\alpha$  in promotion and progression of cancer. *Cancer Metastasis Rev.* 25, 409–416
- Digel, W., Schoniger, W., Stefanic, M., Janssen, H., Buck, C., Schmid, M., Raghavachar, A. and Porzolt, F. (1990) Receptors for tumor necrosis factor on neoplastic B cells from chronic lymphocytic leukemia are expressed *in vitro* but not *in vivo*. *Blood* 76, 1607–1613
- Liu, R. Y., Fan, C., Mitchell, S., Chen, Q., Wu, J. and Zuckerman, K. S. (1998) The role of type I and type II tumor necrosis factor (TNF) receptors in the ability of TNF- $\alpha$  to transduce a proliferative signal in the human megakaryoblastic leukemic cell line Mo7e. *Cancer Res.* 58, 2217–2223
- Tsukasaki, K., Miller, C. W., Kubota, T., Takeuchi, S., Fujimoto, T., Ikeda, S., Tomonaga, M. and Koeffler, H. P. (2001) Tumor necrosis factor  $\alpha$  polymorphism associated with increased susceptibility to development of adult T-cell leukemia/lymphoma in human T-lymphotropic virus type 1 carriers. *Cancer Res.* 61, 3770–3774
- Montesano, R., Soulie, P., Eble, J. A. and Carrozzino, F. (2005) Tumour necrosis factor  $\alpha$  confers an invasive, transformed phenotype on mammary epithelial cells. *J. Cell Sci.* 118, 3487–3500
- Kalthoff, H., Roeder, C., Gieseck, J., Humburg, I. and Schmielgel, W. (1993) Inverse regulation of human ERBB2 and epidermal growth factor receptors by tumor necrosis factor  $\alpha$ . *Proc. Natl. Acad. Sci. U.S.A.* 90, 8972–8976
- Schmielgel, W., Roeder, C., Schmielau, J., Rodeck, U. and Kalthoff, H. (1993) Tumor necrosis factor  $\alpha$  induces the expression of transforming growth factor  $\alpha$  and the epidermal growth factor receptor in human pancreatic cancer cells. *Proc. Natl. Acad. Sci. U.S.A.* 90, 863–867
- Naylor, M. S., Stamp, G. W., Foulkes, W. D., Eccles, D. and Balkwill, F. R. (1993) Tumor necrosis factor and its receptors in human ovarian cancer. Potential role in disease progression. *J. Clin. Invest.* 91, 2194–2206
- Duarte, I., Santos, A., Sousa, H., Catarino, R., Pinto, D., Matos, A., Pereira, D., Moutinho, J., Canedo, P., Machado, J. C. and Medeiros, R. (2005) G-308A TNF- $\alpha$  polymorphism is associated with an increased risk of invasive cervical cancer. *Biochem. Biophys. Res. Commun.* 334, 588–592
- Aggarwal, B. B., Schwarz, L., Hogan, M. E. and Rando, R. F. (1996) Triple helix-forming oligodeoxynucleotides targeted to the human tumor necrosis factor (TNF) gene inhibit TNF production and block the TNF-dependent growth of human glioblastoma tumor cells. *Cancer Res.* 56, 5156–5164



- 26 Nabors, L. B., Suswam, E., Huang, Y., Yang, X., Johnson, M. J. and King, P. H. (2003) Tumor necrosis factor  $\alpha$  induces angiogenic factor up-regulation in malignant glioma cells: a role for RNA stabilization and HuR. *Cancer Res.* 63, 4181–4187
- 27 Pikarsky, E., Porat, R. M., Stein, I., Abramovitch, R., Amit, S., Kasem, S., Galkovich-Pyest, E., Urieli-Shoval, S., Galun, E. and Ben-Neriah, Y. (2004) NF- $\kappa$ B functions as a tumour promoter in inflammation-associated cancer. *Nature* 431, 461–466
- 28 Orosz, P., Echtenacher, B., Falk, W., Ruschoff, J., Weber, D. and Mannel, D. N. (1993) Enhancement of experimental metastasis by tumor necrosis factor. *J. Exp. Med.* 177, 1391–1398
- 29 Ferrajoli, A., Keating, M. J., Manshour, T., Giles, F. J., Dey, A., Estrov, Z., Koller, C. A., Kurzrock, R., Thomas, D. A., Faderl, S. et al. (2002) The clinical significance of tumor necrosis factor- $\alpha$  plasma level in patients having chronic lymphocytic leukemia. *Blood* 100, 1215–1219
- 30 Wolf, J. S., Chen, Z., Dong, G., Sunwoo, J. B., Bancroft, C. C., Capo, D. E., Yeh, N. T., Mukaida, N. and Van Waes, C. (2001) IL (interleukin)-1 $\alpha$  promotes nuclear factor- $\kappa$ B and AP-1-induced IL-8 expression, cell survival, and proliferation in head and neck squamous cell carcinomas. *Clin. Cancer Res.* 7, 1812–1820
- 31 Lust, J. A., Lacy, M. Q., Zeldenrust, S. R., Dispenzieri, A., Gertz, M. A., Witzig, T. E., Kumar, S., Hayman, S. R., Russell, S. J., Buadi, F. K. et al. (2009) Induction of a chronic disease state in patients with smoldering or indolent multiple myeloma by targeting interleukin 1 $\beta$ -induced interleukin 6 production and the myeloma proliferative component. *Mayo Clin. Proc.* 84, 114–122
- 32 Jung, Y. J., Isaacs, J. S., Lee, S., Trepel, J. and Neckers, L. (2003) IL-1 $\beta$ -mediated up-regulation of HIF-1 $\alpha$  via an NF- $\kappa$ B/COX-2 pathway identifies HIF-1 as a critical link between inflammation and oncogenesis. *FASEB J.* 17, 2115–2117
- 33 Abramov, Y., Anteby, S. O., Fasoulitis, S. J. and Barak, V. (2002) The role of inflammatory cytokines in Meigs' syndrome. *Obstet. Gynecol.* 99, 917–919
- 34 Hong, D. S., Angelo, L. S. and Kurzrock, R. (2007) Interleukin-6 and its receptor in cancer: implications for translational therapeutics. *Cancer* 110, 1911–1928
- 35 Naugler, W. E. and Karin, M. (2008) The wolf in sheep's clothing: the role of interleukin-6 in immunity, inflammation and cancer. *Trends Mol. Med.* 14, 109–119
- 36 Lin, W. W. and Karin, M. (2007) A cytokine-mediated link between innate immunity, inflammation, and cancer. *J. Clin. Invest.* 117, 1175–1183
- 37 Angelo, L. S., Talpaz, M. and Kurzrock, R. (2002) Autocrine interleukin-6 production in renal cell carcinoma: evidence for the involvement of p53. *Cancer Res.* 62, 932–940
- 38 Kai, H., Kitadai, Y., Kodama, M., Cho, S., Kuroda, T., Ito, M., Tanaka, S., Ohmoto, Y. and Chayama, K. (2005) Involvement of proinflammatory cytokines IL-1 $\beta$  and IL-6 in progression of human gastric carcinoma. *Anticancer Res.* 25, 709–713
- 39 Klein, B. and Bataille, R. (1992) Cytokine network in human multiple myeloma. *Hematol. Oncol. Clin. North Am.* 6, 273–284
- 40 Chung, Y. C. and Chang, Y. F. (2003) Serum interleukin-6 levels reflect the disease status of colorectal cancer. *J. Surg. Oncol.* 83, 222–226
- 41 Sharma, R., Zucknick, M., London, R., Kacevska, M., Liddle, C. and Clarke, S. J. (2008) Systemic inflammatory response predicts prognosis in patients with advanced-stage colorectal cancer. *Clin. Colorectal Cancer* 7, 331–337
- 42 Waugh, D. J. and Wilson, C. (2008) The interleukin-8 pathway in cancer. *Clin. Cancer Res.* 14, 6735–6741
- 43 Xu, L. and Fidler, I. J. (2000) Acidic pH-induced elevation in interleukin 8 expression by human ovarian carcinoma cells. *Cancer Res.* 60, 4610–4616
- 44 Huang, S., Mills, L., Mian, B., Tellez, C., McCarty, M., Yang, X. D., Gudas, J. M. and Bar-Eli, M. (2002) Fully humanized neutralizing antibodies to interleukin-8 (ABX-IL8) inhibit angiogenesis, tumor growth, and metastasis of human melanoma. *Am. J. Pathol.* 161, 125–134
- 45 Bendre, M. S., Margulies, A. G., Walser, B., Akel, N. S., Bhattacharya, S., Skinner, R. A., Swain, F., Ramani, V., Mohammad, K. S., Wessner, L. L. et al. (2005) Tumor-derived interleukin-8 stimulates osteolysis independent of the receptor activator of nuclear factor- $\kappa$ B ligand pathway. *Cancer Res.* 65, 11001–11009
- 46 Maxwell, P. J., Gallagher, R., Seaton, A., Wilson, C., Scullin, P., Pettigrew, J., Stratford, I. J., Williams, K. J., Johnston, P. G. and Waugh, D. J. (2007) HIF-1 and NF- $\kappa$ B-mediated upregulation of CXCR1 and CXCR2 expression promotes cell survival in hypoxic prostate cancer cells. *Oncogene* 26, 7333–7345
- 47 Asarch, A., Barak, O., Loo, D. S. and Gottlieb, A. B. (2008) Th17 cells: a new therapeutic target in inflammatory dermatoses. *J. Dermatolog. Treat.* 19, 318–326
- 48 Tartour, E., Fossiez, F., Joyeux, I., Galinha, A., Gey, A., Claret, E., Sastre-Garau, X., Couturier, J., Mosseri, V., Vives, V. et al. (1999) Interleukin 17, a T-cell-derived cytokine, promotes tumorigenicity of human cervical tumors in nude mice. *Cancer Res.* 59, 3698–3704
- 49 Numasaki, M., Fukushi, J., Ono, M., Narula, S. K., Zavodny, P. J., Kudo, T., Robbins, P. D., Tahara, H. and Lotze, M. T. (2003) Interleukin-17 promotes angiogenesis and tumor growth. *Blood* 101, 2620–2627
- 50 Numasaki, M., Watanabe, M., Suzuki, T., Takahashi, H., Nakamura, A., McAllister, F., Hishinuma, T., Goto, J., Lotze, M. T., Kolls, J. K. and Sasaki, H. (2005) IL-17 enhances the net angiogenic activity and *in vivo* growth of human non-small cell lung cancer in SCID mice through promoting CXCR-2-dependent angiogenesis. *J. Immunol.* 175, 6177–6189
- 51 Wang, D., Dubois, R. N. and Richmond, A. (2009) The role of chemokines in intestinal inflammation and cancer. *Curr. Opin. Pharmacol.* 9, 688–696
- 52 Lu, H., Ouyang, W. and Huang, C. (2006) Inflammation, a key event in cancer development. *Mol. Cancer Res.* 4, 221–233
- 53 Kollmar, O., Rupertus, K., Scheuer, C., Junker, B., Tilton, B., Schilling, M. K. and Menger, M. D. (2007) Stromal cell-derived factor-1 promotes cell migration and tumor growth of colorectal metastasis. *Neoplasia* 9, 862–870
- 54 Owen, J. D., Strieter, R., Burdick, M., Haghnegahdar, H., Nanney, L., Shattuck-Brandt, R. and Richmond, A. (1997) Enhanced tumor-forming capacity for immortalized melanocytes expressing melanoma growth stimulatory activity/growth-regulated cytokine  $\beta$  and  $\gamma$  proteins. *Int. J. Cancer* 73, 94–103
- 55 Yuecheng, Y. and Xiaoyan, X. (2007) Stromal-cell derived factor-1 regulates epithelial ovarian cancer cell invasion by activating matrix metalloproteinase-9 and matrix metalloproteinase-2. *Eur. J. Cancer Prev.* 16, 430–435
- 56 Muller, A., Homey, B., Soto, H., Ge, N., Catron, D., Buchanan, M. E., McClanahan, T., Murphy, E., Yuan, W., Wagner, S. N. et al. (2001) Involvement of chemokine receptors in breast cancer metastasis. *Nature* 410, 50–56
- 57 Johrer, K., Zelle-Rieser, C., Perathoner, A., Moser, P., Hager, M., Ramoner, R., Gander, H., Holtl, L., Bartsch, G., Greil, R. and Thurnher, M. (2005) Up-regulation of functional chemokine receptor CCR3 in human renal cell carcinoma. *Clin. Cancer Res.* 11, 2459–2465
- 58 Schwarze, S. R., Luo, J., Isaacs, W. B. and Jarrard, D. F. (2005) Modulation of CXCL14 (BRAX) expression in prostate cancer. *Prostate* 64, 67–74

- 59 Kim, S. Y., Lee, C. H., Midura, B. V., Yeung, C., Mendoza, A., Hong, S. H., Ren, L., Wong, D., Korz, W., Merzouk, A. et al. (2008) Inhibition of the CXCR4/CXCL12 chemokine pathway reduces the development of murine pulmonary metastases. *Clin. Exp. Metastasis* 25, 201–211
- 60 Weinberg, R. A. (1994) Oncogenes and tumor suppressor genes. *CA Cancer J. Clin.* 44, 160–170
- 61 Croce, C. M. (2008) Oncogenes and cancer. *N. Engl. J. Med.* 358, 502–511
- 62 Borrello, M. G., Degl'Innocenti, D. and Pierotti, M. A. (2008) Inflammation and cancer: the oncogene-driven connection. *Cancer Lett.* 267, 262–270
- 63 Grivennikov, S. I. and Karin, M. (2010) Inflammation and oncogenesis: a vicious connection. *Curr. Opin. Genet. Dev.* 20, 65–71
- 64 Liu, J., Yang, G., Thompson-Lanza, J. A., Glassman, A., Hayes, K., Patterson, A., Marquez, R. T., Auersperg, N., Yu, Y., Hahn, W. C. et al. (2004) A genetically defined model for human ovarian cancer. *Cancer Res.* 64, 1655–1663
- 65 Yoneda, J., Kuniyasu, H., Crispen, M. A., Price, J. E., Bucana, C. D. and Fidler, I. J. (1998) Expression of angiogenesis-related genes and progression of human ovarian carcinomas in nude mice. *J. Natl. Cancer Inst.* 90, 447–454
- 66 Ancrile, B., Lim, K. H. and Counter, C. M. (2007) Oncogenic Ras-induced secretion of IL6 is required for tumorigenesis. *Genes Dev.* 21, 1714–1719
- 67 Meyer, N. and Penn, L. Z. (2008) Reflecting on 25 years with MYC. *Nat. Rev. Cancer* 8, 976–990
- 68 Shchors, K., Shchors, E., Rostker, F., Lawlor, E. R., Brown-Swigart, L. and Evan, G. I. (2006) The Myc-dependent angiogenic switch in tumors is mediated by interleukin 1 $\beta$ . *Genes Dev.* 20, 2527–2538
- 69 Soucek, L., Lawlor, E. R., Soto, D., Shchors, K., Swigart, L. B. and Evan, G. I. (2007) Mast cells are required for angiogenesis and macroscopic expansion of Myc-induced pancreatic islet tumors. *Nat. Med.* 13, 1211–1218
- 70 Arighi, E., Borrello, M. G. and Sariola, H. (2005) RET tyrosine kinase signaling in development and cancer. *Cytokine Growth Factor Rev.* 16, 441–467
- 71 Hwang, E. S. and Bowen, P. E. (2007) DNA damage, a biomarker of carcinogenesis: its measurement and modulation by diet and environment. *Crit. Rev. Food Sci. Nutr.* 47, 27–50
- 72 Schetter, A. J., Heegaard, N. H. and Harris, C. C. (2010) Inflammation and cancer: interweaving microRNA, free radical, cytokine and p53 pathways. *Carcinogenesis* 31, 37–49
- 73 Klaunig, J. E., Kamendulis, L. M. and Hocevar, B. A. (2010) Oxidative stress and oxidative damage in carcinogenesis. *Toxicol. Pathol.* 38, 96–109
- 74 Babior, B. M. (2000) Phagocytes and oxidative stress. *Am. J. Med.* 109, 33–44
- 75 Closa, D. and Folch-Puy, E. (2004) Oxygen free radicals and the systemic inflammatory response. *IUBMB Life* 56, 185–191
- 76 Kwak, M. K. and Kensler, T. W. (2010) Targeting NRF2 signaling for cancer chemoprevention. *Toxicol. Appl. Pharmacol.* 244, 66–76
- 77 Marnett, L. J. (2000) Oxyradicals and DNA damage. *Carcinogenesis* 21, 361–370
- 78 Shen, H. M. and Ong, C. N. (1996) Mutations of the p53 tumor suppressor gene and ras oncogenes in aflatoxin hepatocarcinogenesis. *Mutat. Res.* 366, 23–44
- 79 Nishigori, C., Hattori, Y. and Toyokuni, S. (2004) Role of reactive oxygen species in skin carcinogenesis. *Antioxid. Redox Signal.* 6, 561–570
- 80 Aggarwal, B. B. and Shishodia, S. (2006) Molecular targets of dietary agents for prevention and therapy of cancer. *Biochem. Pharmacol.* 71, 1397–1421
- 81 Aggarwal, B. B., Van Kuiken, M. E., Iyer, L. H., Harikumar, K. B. and Sung, B. (2009) Molecular targets of nutraceuticals derived from dietary spices: potential role in suppression of inflammation and tumorigenesis. *Exp. Biol. Med.* (Maywood) 234, 825–849
- 82 Aggarwal, B. B., Kunnumakkara, A. B., Harikumar, K. B., Tharakan, S. T., Sung, B. and Anand, P. (2008) Potential of spice-derived phytochemicals for cancer prevention. *Planta Med.* 74, 1560–1569
- 83 Surh, Y. J. and Kundu, J. K. (2007) Cancer preventive phytochemicals as speed breakers in inflammatory signaling involved in aberrant COX-2 expression. *Curr. Cancer Drug Targets* 7, 447–458
- 84 Einspahr, J. G., Krouse, R. S., Yochim, J. M., Danenberg, P. V., Danenberg, K. D., Bhattacharyya, A. K., Martinez, M. E. and Alberts, D. S. (2003) Association between cyclooxygenase expression and colorectal adenoma characteristics. *Cancer Res.* 63, 3891–3893
- 85 Oshima, M., Dinchuk, J. E., Kargman, S. L., Oshima, H., Hancock, B., Kwong, E., Trzaskos, J. M., Evans, J. F. and Taketo, M. M. (1996) Suppression of intestinal polyposis in Apc  $\Delta$ 716 knockout mice by inhibition of cyclooxygenase 2 (COX-2). *Cell* 87, 803–809
- 86 Oshima, M., Murai, N., Kargman, S., Arguello, M., Luk, P., Kwong, E., Taketo, M. M. and Evans, J. F. (2001) Chemoprevention of intestinal polyposis in the Apc $\Delta$ 716 mouse by rofecoxib, a specific cyclooxygenase-2 inhibitor. *Cancer Res.* 61, 1733–1740
- 87 Liu, W., Reinmuth, N., Stoeltzing, O., Parikh, A. A., Tellez, C., Williams, S., Jung, Y. D., Fan, F., Takeda, A., Akagi, M. et al. (2003) Cyclooxygenase-2 is up-regulated by interleukin-1  $\beta$  in human colorectal cancer cells via multiple signaling pathways. *Cancer Res.* 63, 3632–3636
- 88 Pold, M., Zhu, L. X., Sharma, S., Burdick, M. D., Lin, Y., Lee, P. P., Pold, A., Luo, J., Krysan, K., Dohadwala, M. et al. (2004) Cyclooxygenase-2-dependent expression of angiogenic CXC chemokines ENA-78/CXC ligand (CXCL) 5 and interleukin-8/CXCL8 in human non-small cell lung cancer. *Cancer Res.* 64, 1853–1860
- 89 Hashimoto, N., Inayama, M., Fujishima, M. and Shiozaki, H. (2007) Clinicopathologic significance of expression of cyclooxygenase-2 in human esophageal squamous cell carcinoma. *Hepatogastroenterology* 54, 758–760
- 90 Shono, T., Tofilon, P. J., Bruner, J. M., Owlabi, O. and Lang, F. F. (2001) Cyclooxygenase-2 expression in human gliomas: prognostic significance and molecular correlations. *Cancer Res.* 61, 4375–4381
- 91 Kim, Y. B., Kim, G. E., Cho, N. H., Pyo, H. R., Shim, S. J., Chang, S. K., Park, H. C., Suh, C. O., Park, T. K. and Kim, B. S. (2002) Overexpression of cyclooxygenase-2 is associated with a poor prognosis in patients with squamous cell carcinoma of the uterine cervix treated with radiation and concurrent chemotherapy. *Cancer* 95, 531–539
- 92 Cetin, M., Buyukberber, S., Demir, M., Sari, I., Deniz, K., Eser, B., Altuntas, F., Camci, C., Ozturk, A., Turgut, B. et al. (2005) Overexpression of cyclooxygenase-2 in multiple myeloma: association with reduced survival. *Am. J. Hematol.* 80, 169–173
- 93 Pidgeon, G. P., Lysaght, J., Krishnamoorthy, S., Reynolds, J. V., O'Byrne, K., Nie, D. and Honn, K. V. (2007) Lipoxigenase metabolism: roles in tumor progression and survival. *Cancer Metastasis Rev.* 26, 503–524
- 94 Cuendet, M. and Pezzuto, J. M. (2000) The role of cyclooxygenase and lipoxigenase in cancer chemoprevention. *Drug Metabol. Drug Interact.* 17, 109–157
- 95 Shureiqi, I. and Lippman, S. M. (2001) Lipoxigenase modulation to reverse carcinogenesis. *Cancer Res.* 61, 6307–6312
- 96 Boado, R. J., Pardridge, W. M., Vinters, H. V. and Black, K. L. (1992) Differential expression of arachidonate 5-lipoxygenase transcripts in human brain tumors: evidence for the expression of a multitranscript family. *Proc. Natl. Acad. Sci. U.S.A.* 89, 9044–9048



- 97 Ye, Y. N., Liu, E. S., Shin, V. Y., Wu, W. K. and Cho, C. H. (2004) Contributory role of 5-lipoxygenase and its association with angiogenesis in the promotion of inflammation-associated colonic tumorigenesis by cigarette smoking. *Toxicology* 203, 179–188
- 98 Hennig, R., Grippo, P., Ding, X. Z., Rao, S. M., Buchler, M. W., Friess, H., Talamonti, M. S., Bell, R. H. and Adrian, T. E. (2005) 5-Lipoxygenase, a marker for early pancreatic intraepithelial neoplastic lesions. *Cancer Res.* 65, 6011–6016
- 99 Jiang, W. G., Douglas-Jones, A. G. and Mansel, R. E. (2006) Aberrant expression of 5-lipoxygenase-activating protein (5-LOXAP) has prognostic and survival significance in patients with breast cancer. *Prostaglandins Leukot. Essent. Fatty Acids* 74, 125–134
- 100 Faronato, M., Muzzonigro, G., Milanese, G., Menna, C., Bonfigli, A. R., Catalano, A. and Procopio, A. (2007) Increased expression of 5-lipoxygenase is common in clear cell renal cell carcinoma. *Histol. Histopathol.* 22, 1109–1118
- 101 Cruz-Munoz, W. and Khokha, R. (2008) The role of tissue inhibitors of metalloproteinases in tumorigenesis and metastasis. *Crit. Rev. Clin. Lab. Sci.* 45, 291–338
- 102 Thiery, J. P. (2002) Epithelial-mesenchymal transitions in tumour progression. *Nat. Rev. Cancer* 2, 442–454
- 103 Bergers, G., Brekken, R., McMahon, G., Vu, T. H., Itoh, T., Tamaki, K., Tanzawa, K., Thorpe, P., Itohara, S., Werb, Z. and Hanahan, D. (2000) Matrix metalloproteinase-9 triggers the angiogenic switch during carcinogenesis. *Nat. Cell Biol.* 2, 737–744
- 104 Coussens, L. M., Tinkle, C. L., Hanahan, D. and Werb, Z. (2000) MMP-9 supplied by bone marrow-derived cells contributes to skin carcinogenesis. *Cell* 103, 481–490
- 105 van't Veer, L. J., Dai, H., van de Vijver, M. J., He, Y. D., Hart, A. A., Mao, M., Peterse, H. L., van der Kooy, K., Marton, M. J., Witteveen, A. T. et al. (2002) Gene expression profiling predicts clinical outcome of breast cancer. *Nature* 415, 530–536
- 106 Minn, A. J., Gupta, G. P., Siegel, P. M., Bos, P. D., Shu, W., Giri, D. D., Viale, A., Olshen, A. B., Gerald, W. L. and Massague, J. (2005) Genes that mediate breast cancer metastasis to lung. *Nature* 436, 518–524
- 107 Chen, X. L., Wang, L. C., Zhang, W. G., Chen, X. Y. and Sun, Z. M. (2008) Correlations of S100A4 and MMP9 expressions to infiltration, metastasis and prognosis of non-small cell lung cancer. *Nan Fang Yi Ke Da Xue Xue Bao* 28, 1254–1258
- 108 Hu, Z. L., Wen, J. F., Shen, M. and Liu, Y. (2006) Expressions of TGIF, MMP9 and VEGF proteins and their clinicopathological relationship in gastric cancer. *Zhong Nan Da Xue Xue Bao Yi Xue Ban* 31, 70–74
- 109 Gu, Z. D., Li, J. Y., Li, M., Gu, J., Shi, X. T., Ke, Y. and Chen, K. N. (2005) Matrix metalloproteinases expression correlates with survival in patients with esophageal squamous cell carcinoma. *Am. J. Gastroenterol.* 100, 1835–1843
- 110 Sakata, K., Satoh, M., Someya, M., Asanuma, H., Nagakura, H., Oouchi, A., Nakata, K., Kogawa, K., Koito, K., Hareyama, M. and Himi, T. (2004) Expression of matrix metalloproteinase 9 is a prognostic factor in patients with non-Hodgkin lymphoma. *Cancer* 100, 356–365
- 111 Karin, M. (2006) Nuclear factor- $\kappa$ B in cancer development and progression. *Nature* 441, 431–436
- 112 Sethi, G. and Tergaonkar, V. (2009) Potential pharmacological control of the NF- $\kappa$ B pathway. *Trends Pharmacol. Sci.* 30, 313–321
- 113 Sethi, G., Sung, B. and Aggarwal, B. B. (2008) Nuclear factor- $\kappa$ B activation: from bench to bedside. *Exp. Biol. Med.* (Maywood) 233, 21–31
- 114 Tergaonkar, V., Correa, R. G., Ikawa, M. and Verma, I. M. (2005) Distinct roles of I $\kappa$ B proteins in regulating constitutive NF- $\kappa$ B activity. *Nat. Cell Biol.* 7, 921–923
- 115 Karin, M. (2006) NF- $\kappa$ B and cancer: mechanisms and targets. *Mol. Carcinog.* 45, 355–361
- 116 Vallabhapurapu, S. and Karin, M. (2009) Regulation and function of NF- $\kappa$ B transcription factors in the immune system. *Annu. Rev. Immunol.* 27, 693–733
- 117 Shen, H. M. and Tergaonkar, V. (2009) NF- $\kappa$ B signaling in carcinogenesis and as a potential molecular target for cancer therapy. *Apoptosis* 14, 348–363
- 118 Ahn, K. S., Sethi, G. and Aggarwal, B. B. (2007) Nuclear factor- $\kappa$ B: from clone to clinic. *Curr. Mol. Med.* 7, 619–637
- 119 Ahn, K. S. and Aggarwal, B. B. (2005) Transcription factor NF- $\kappa$ B: a sensor for smoke and stress signals. *Ann. N.Y. Acad. Sci.* 1056, 218–233
- 120 Biswas, S. K., Bist, P., Dhillon, M. K., Kajiji, T., Del Fresno, C., Yamamoto, M., Lopez-Collazo, E., Akira, S. and Tergaonkar, V. (2007) Role for MyD88-independent, TRIF pathway in lipid A/TLR4-induced endotoxin tolerance. *J. Immunol.* 179, 4083–4092
- 121 Lee, K. G., Xu, S., Wong, E. T., Tergaonkar, V. and Lam, K. P. (2008) Bruton's tyrosine kinase separately regulates NF- $\kappa$ B p65RelA activation and cytokine interleukin (IL)-10/IL-12 production in TLR9-stimulated B Cells. *J. Biol. Chem.* 283, 11189–11198
- 122 Wong, E. T. and Tergaonkar, V. (2009) Roles of NF- $\kappa$ B in health and disease: mechanisms and therapeutic potential. *Clin. Sci.* 116, 451–465
- 123 Mantovani, A. (2010) Molecular pathways linking inflammation and cancer. *Curr. Mol. Med.* 10, 369–373
- 124 Li, F. and Sethi, G. (2010) Targeting transcription factor NF- $\kappa$ B to overcome chemoresistance and radioresistance in cancer therapy. *Biochim. Biophys. Acta* 1805, 167–180
- 125 Baud, V. and Karin, M. (2009) Is NF- $\kappa$ B a good target for cancer therapy? Hopes and pitfalls. *Nat. Rev. Drug Discov.* 8, 33–40
- 126 Greten, F. R., Eckmann, L., Greten, T. F., Park, J. M., Li, Z. W., Egan, L. J., Kagnoff, M. F. and Karin, M. (2004) IKK $\beta$  links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. *Cell* 118, 285–296
- 127 Naugler, W. E. and Karin, M. (2008) NF- $\kappa$ B and cancer-identifying targets and mechanisms. *Curr. Opin. Genet. Dev.* 18, 19–26
- 128 Karin, M. (2008) The I $\kappa$ B kinase – a bridge between inflammation and cancer. *Cell Res.* 18, 334–342
- 129 Chew, J., Biswas, S., Shreeram, S., Humaidi, M., Wong, E. T., Dhillon, M. K., Teo, H., Hazra, A., Fang, C. C., Lopez-Collazo, E. et al. (2009) WIP1 phosphatase is a negative regulator of NF- $\kappa$ B signalling. *Nat. Cell Biol.* 11, 659–666
- 130 Tergaonkar, V., Pando, M., Vafa, O., Wahl, G. and Verma, I. (2002) p53 stabilization is decreased upon NF- $\kappa$ B activation: a role for NF- $\kappa$ B in acquisition of resistance to chemotherapy. *Cancer Cell* 1, 493–503
- 131 Tergaonkar, V. and Perkins, N. D. (2007) p53 and NF- $\kappa$ B crosstalk: IKK $\alpha$  tips the balance. *Mol. Cell* 26, 158–159
- 132 Xia, Y., Padre, R. C., De Mendoza, T. H., Bottero, V., Tergaonkar, V. B. and Verma, I. M. (2009) Phosphorylation of p53 by I $\kappa$ B kinase 2 promotes its degradation by  $\beta$ -TrCP. *Proc. Natl. Acad. Sci. U.S.A.* 106, 2629–2634
- 133 Irelan, J. T., Murphy, T. J., DeJesus, P. D., Teo, H., Xu, D., Gomez-Ferreria, M. A., Zhou, Y., Miraglia, L. J., Rines, D. R., Verma, I. M. et al. (2007) A role for I $\kappa$ B kinase 2 in bipolar spindle assembly. *Proc. Natl. Acad. Sci. U.S.A.* 104, 16940–16945
- 134 Dey, A., Wong, E. T., Bist, P., Tergaonkar, V. and Lane, D. P. (2007) Nutlin-3 inhibits the NF- $\kappa$ B pathway in a p53-dependent manner: implications in lung cancer therapy. *Cell Cycle* 6, 2178–2185
- 135 Dey, A., Wong, E. T., Cheok, C. F., Tergaonkar, V. and Lane, D. P. (2008) R-Roscovitine simultaneously targets both the p53 and NF- $\kappa$ B pathways and causes potentiation of apoptosis: implications in cancer therapy. *Cell Death Differ.* 15, 263–273
- 136 Dey, A., Tergaonkar, V. and Lane, D. P. (2008) Double-edged swords as cancer therapeutics: simultaneously targeting p53 and NF- $\kappa$ B pathways. *Nat. Rev. Drug Discov.* 7, 1031–1040

- 137 Mantovani, A. and Sica, A. (2010) Macrophages, innate immunity and cancer: balance, tolerance, and diversity. *Curr. Opin. Immunol.* 22, 231–237
- 138 Uzzo, R. G., Clark, P. E., Rayman, P., Bloom, T., Rybicki, L., Novick, A. C., Bukowski, R. M. and Finke, J. H. (1999) Alterations in NF $\kappa$ B activation in T lymphocytes of patients with renal cell carcinoma. *J. Natl. Cancer Inst.* 91, 718–721
- 139 Balkwill, F., Charles, K. A. and Mantovani, A. (2005) Smoldering and polarized inflammation in the initiation and promotion of malignant disease. *Cancer Cell* 7, 211–217
- 140 Biswas, S. K., Gangi, L., Paul, S., Schioppa, T., Saccani, A., Sironi, M., Bottazzi, B., Doni, A., Vincenzo, B., Pasqualini, F. et al. (2006) A distinct and unique transcriptional program expressed by tumor-associated macrophages (defective NF $\kappa$ B and enhanced IRF-3/STAT1 activation). *Blood* 107, 2112–2122
- 141 Sica, A., Larghi, P., Mancino, A., Rubino, L., Porta, C., Totaro, M. G., Rimoldi, M., Biswas, S. K., Allavena, P. and Mantovani, A. (2008) Macrophage polarization in tumour progression. *Semin. Cancer Biol.* 18, 349–355
- 142 Saccani, A., Schioppa, T., Porta, C., Biswas, S. K., Nebuloni, M., Vago, L., Bottazzi, B., Colombo, M. P., Mantovani, A. and Sica, A. (2006) p50 nuclear factor- $\kappa$ B overexpression in tumor-associated macrophages inhibits M1 inflammatory responses and antitumor resistance. *Cancer Res.* 66, 11432–11440
- 143 Nickoloff, B. J., Ben-Neriah, Y. and Pikarsky, E. (2005) Inflammation and cancer: is the link as simple as we think? *J. Invest. Dermatol.* 124, x–xiv
- 144 Mantovani, A., Bottazzi, B., Colotta, F., Sozzani, S. and Ruco, L. (1992) The origin and function of tumor-associated macrophages. *Immunol. Today* 13, 265–270
- 145 Hagemann, T., Lawrence, T., McNeish, I., Charles, K. A., Kulbe, H., Thompson, R. G., Robinson, S. C. and Balkwill, F. R. (2008) “Re-educating” tumor-associated macrophages by targeting NF $\kappa$ B. *J. Exp. Med.* 205, 1261–1268
- 146 Ihle, J. N. (1996) STATs: signal transducers and activators of transcription. *Cell* 84, 331–334
- 147 Yu, H. and Jove, R. (2004) The STATs of cancer – new molecular targets come of age. *Nat. Rev. Cancer* 4, 97–105
- 148 Bowman, T., Garcia, R., Turkson, J. and Jove, R. (2000) STATs in oncogenesis. *Oncogene* 19, 2474–2488
- 149 Yu, H., Pardoll, D. and Jove, R. (2009) STATs in cancer inflammation and immunity: a leading role for STAT3. *Nat. Rev. Cancer* 9, 798–809
- 150 Aggarwal, B. B., Kunnumakkara, A. B., Harikumar, K. B., Gupta, S. R., Tharakan, S. T., Koca, C., Dey, S. and Sung, B. (2009) Signal transducer and activator of transcription-3, inflammation, and cancer: how intimate is the relationship? *Ann. N.Y. Acad. Sci.* 1171, 59–76
- 151 Turkson, J. and Jove, R. (2000) STAT proteins: novel molecular targets for cancer drug discovery. *Oncogene* 19, 6613–6626
- 152 Aggarwal, B. B., Sethi, G., Ahn, K. S., Sandur, S. K., Pandey, M. K., Kunnumakkara, A. B., Sung, B. and Ichikawa, H. (2006) Targeting signal-transducer-and-activator-of-transcription-3 for prevention and therapy of cancer: modern target but ancient solution. *Ann. N.Y. Acad. Sci.* 1091, 151–169
- 153 Rebouissou, S., Amessou, M., Couchy, G., Poussin, K., Imbeaud, S., Pilati, C., Izard, T., Balabaud, C., Bioulac-Sage, P. and Zucman-Rossi, J. (2009) Frequent in-frame somatic deletions activate gp130 in inflammatory hepatocellular tumours. *Nature* 457, 200–204
- 154 Li, Y., Du, H., Qin, Y., Roberts, J., Cummings, O. W. and Yan, C. (2007) Activation of the signal transducers and activators of the transcription 3 pathway in alveolar epithelial cells induces inflammation and adenocarcinomas in mouse lung. *Cancer Res.* 67, 8494–8503
- 155 Ernst, M., Najdovska, M., Grail, D., Lundgren-May, T., Buchert, M., Tye, H., Matthews, V. B., Armes, J., Bhathal, P. S., Hughes, N. R. et al. (2008) STAT3 and STAT1 mediate IL-11-dependent and inflammation-associated gastric tumorigenesis in gp130 receptor mutant mice. *J. Clin. Invest.* 118, 1727–1738
- 156 Bronte-Tinkew, D. M., Terebiznik, M., Franco, A., Ang, M., Ahn, D., Mimuro, H., Sasakawa, C., Ropeleski, M. J., Peek, Jr, R. M. and Jones, N. L. (2009) *Helicobacter pylori* cytotoxin-associated gene A activates the signal transducer and activator of transcription 3 pathway *in vitro* and *in vivo*. *Cancer Res.* 69, 632–639
- 157 Sano, S., Chan, K. S., Kira, M., Kataoka, K., Takagi, S., Tarutani, M., Itami, S., Kiguchi, K., Yokoi, M., Sugawara, K. et al. (2005) Signal transducer and activator of transcription 3 is a key regulator of keratinocyte survival and proliferation following UV irradiation. *Cancer Res.* 65, 5720–5729
- 158 Arredondo, J., Chernyavsky, A. I., Jolkovsky, D. L., Pinkerton, K. E. and Grando, S. A. (2006) Receptor-mediated tobacco toxicity: cooperation of the Ras/Raf-1/MEK1/ERK and JAK-2/STAT-3 pathways downstream of  $\alpha$ 7 nicotinic receptor in oral keratinocytes. *FASEB J.* 20, 2093–2101
- 159 Grivennikov, S. I. and Karin, M. (2010) Dangerous liaisons: STAT3 and NF $\kappa$ B collaboration and crosstalk in cancer. *Cytokine Growth Factor Rev.* 21, 11–19
- 160 Dauer, D. J., Ferraro, B., Song, L., Yu, B., Mora, L., Buettner, R., Enkemann, S., Jove, R. and Haura, E. B. (2005) Stat3 regulates genes common to both wound healing and cancer. *Oncogene* 24, 3397–3408
- 161 Park, E. J., Lee, J. H., Yu, G. Y., He, G., Ali, S. R., Holzer, R. G., Osterreicher, C. H., Takahashi, H. and Karin, M. (2010) Dietary and genetic obesity promote liver inflammation and tumorigenesis by enhancing IL-6 and TNF expression. *Cell* 140, 197–208
- 162 Angel, P. and Karin, M. (1991) The role of Jun, Fos and the AP-1 complex in cell-proliferation and transformation. *Biochim. Biophys. Acta* 1072, 129–157
- 163 Eferl, R. and Wagner, E. F. (2003) AP-1: a double-edged sword in tumorigenesis. *Nat. Rev. Cancer* 3, 859–868
- 164 Shaulian, E. (2010) AP-1 – the Jun proteins: oncogenes or tumor suppressors in disguise? *Cell Signal.* 22, 894–899
- 165 Young, M. R., Yang, H. S. and Colburn, N. H. (2003) Promising molecular targets for cancer prevention: AP-1, NF $\kappa$ B and Pcd4. *Trends Mol. Med.* 9, 36–41
- 166 Chang, L. and Karin, M. (2001) Mammalian MAP kinase signalling cascades. *Nature* 410, 37–40
- 167 Matthews, C. P., Colburn, N. H. and Young, M. R. (2007) AP-1 a target for cancer prevention. *Curr. Cancer Drug Targets* 7, 317–324
- 168 Ozanne, B. W., Spence, H. J., McGarry, L. C. and Hennigan, R. F. (2007) Transcription factors control invasion: AP-1 the first among equals. *Oncogene* 26, 1–10
- 169 Bos, T. J., Monteclaro, F. S., Mitsunobu, F., Ball, Jr, A. R., Chang, C. H., Nishimura, T. and Vogt, P. K. (1990) Efficient transformation of chicken embryo fibroblasts by c-Jun requires structural modification in coding and noncoding sequences. *Genes Dev.* 4, 1677–1687
- 170 Verde, P., Casalino, L., Talotta, F., Yaniv, M. and Weitzman, J. B. (2007) Deciphering AP-1 function in tumorigenesis: fraternizing on target promoters. *Cell Cycle* 6, 2633–2639
- 171 Dong, Z., Crawford, H. C., Lavrovsky, V., Taub, D., Watts, R., Matrisian, L. M. and Colburn, N. H. (1997) A dominant negative mutant of jun blocking 12-O-tetradecanoylphorbol-13-acetate-induced invasion in mouse keratinocytes. *Mol. Carcinog.* 19, 204–212
- 172 Bos, T. J., Margiotta, P., Bush, L. and Wasilenko, W. (1999) Enhanced cell motility and invasion of chicken embryo fibroblasts in response to Jun over-expression. *Int. J. Cancer* 81, 404–410
- 173 Briggs, J., Chamboredon, S., Castellazzi, M., Kerry, J. A. and Bos, T. J. (2002) Transcriptional upregulation of SPARC, in response to c-Jun overexpression, contributes to increased motility and invasion of MCF7 breast cancer cells. *Oncogene* 21, 7077–7091





- 174 Cohen, S. B., Waha, A., Gelman, I. H. and Vogt, P. K. (2001) Expression of a down-regulated target, SSeCKS, reverses v-Jun-induced transformation of 10T1/2 murine fibroblasts. *Oncogene* 20, 141–146
- 175 Black, E. J., Clair, T., Delrow, J., Neiman, P. and Gillespie, D. A. (2004) Microarray analysis identifies autotaxin, a tumour cell motility and angiogenic factor with lysophospholipase D activity, as a specific target of cell transformation by v-Jun. *Oncogene* 23, 2357–2366
- 176 Eferl, R., Ricci, R., Kenner, L., Zenz, R., David, J. P., Rath, M. and Wagner, E. F. (2003) Liver tumor development. c-Jun antagonizes the proapoptotic activity of p53. *Cell* 112, 181–192
- 177 Young, M. R., Li, J. J., Rincon, M., Flavell, R. A., Sathyanarayana, B. K., Hunziker, R. and Colburn, N. (1999) Transgenic mice demonstrate AP-1 (activator protein-1) transactivation is required for tumor promotion. *Proc. Natl. Acad. Sci. U.S.A.* 96, 9827–9832
- 178 Wang, Z. Q., Grigoriadis, A. E., Mohle-Steinlein, U. and Wagner, E. F. (1991) A novel target cell for c-fos-induced oncogenesis: development of chondrogenic tumours in embryonic stem cell chimeras. *EMBO J.* 10, 2437–2450
- 179 Li, J. J., Rhim, J. S., Schlegel, R., Vousden, K. H. and Colburn, N. H. (1998) Expression of dominant negative Jun inhibits elevated AP-1 and NF- $\kappa$ B transactivation and suppresses anchorage independent growth of HPV immortalized human keratinocytes. *Oncogene* 16, 2711–2721
- 180 Guillemin, K. and Krasnow, M. A. (1997) The hypoxic response: huffing and HIFing. *Cell* 89, 9–12
- 181 Semenza, G. L. (2003) Targeting HIF-1 for cancer therapy. *Nat. Rev. Cancer* 3, 721–732
- 182 Ryan, H. E., Lo, J. and Johnson, R. S. (1998) HIF-1  $\alpha$  is required for solid tumor formation and embryonic vascularization. *EMBO J.* 17, 3005–3015
- 183 Dehne, N. and Brune, B. (2009) HIF-1 in the inflammatory microenvironment. *Exp. Cell Res.* 315, 1791–1797
- 184 Hellwig-Burgel, T., Rutkowski, K., Metzen, E., Fandrey, J. and Jelkmann, W. (1999) Interleukin-1 $\beta$  and tumor necrosis factor- $\alpha$  stimulate DNA binding of hypoxia-inducible factor-1. *Blood* 94, 1561–1567
- 185 Poon, E., Harris, A. L. and Ashcroft, M. (2009) Targeting the hypoxia-inducible factor (HIF) pathway in cancer. *Expert Rev. Mol. Med.* 11, e26
- 186 Koh, M. Y., Spivak-Kroizman, T. R. and Powis, G. (2010) HIF-1 $\alpha$  and cancer therapy. *Recent Results Cancer Res.* 180, 15–34
- 187 Dajee, M., Lazarov, M., Zhang, J. Y., Cai, T., Green, C. L., Russell, A. J., Marinkovich, M. P., Tao, S., Lin, Q., Kubo, Y. and Khavari, P. A. (2003) NF- $\kappa$ B blockade and oncogenic Ras trigger invasive human epidermal neoplasia. *Nature* 421, 639–643
- 188 He, G., Yu, G. Y., Temkin, V., Ogata, H., Kuntzen, C., Sakurai, T., Sieghart, W., Peck-Radosavljevic, M., Leffert, H. L. and Karin, M. (2010) Hepatocyte IKK $\beta$ /NF- $\kappa$ B inhibits tumor promotion and progression by preventing oxidative stress-driven STAT3 activation. *Cancer Cell* 17, 286–297
- 189 Geborek, P., Nitelius, E., Noltorp, S., Petri, H., Jacobsson, L., Larsson, L., Saxne, T. and Leden, I. (2005) Population based studies of biological antirheumatic drug use in southern Sweden: comparison with pharmaceutical sales. *Ann. Rheum. Dis.* 64, 1805–1807
- 190 Wu, Z. H., Wong, E. T., Shi, Y., Niu, J., Chen, Z., Miyamoto, S. and Tergaonkar, V. (2010) ATM- and NEMO-dependent ELKS ubiquitination coordinates TAK1-mediated IKK activation in response to genotoxic stress. *Mol. Cell* 40, 75–86
- 191 Teo, H., Ghosh, S., Luesch, H., Ghosh, A., Wong, E. T., Malik, N., Orth, A., de Jesus, P., Perry, A. S., Oliver, J. D. et al. (2010) Telomere-independent Rap1 is an IKK adaptor and regulates NF- $\kappa$ B-dependent gene expression. *Nat. Cell Biol.* 12, 758–767
- 192 Milner, J. D., Brenchley, J. M., Laurence, A., Freeman, A. F., Hill, B. J., Elias, K. M., Kanno, Y., Spalding, C., Elloumi, H. Z., Paulson, M. L. et al. (2008) Impaired T(H)17 cell differentiation in subjects with autosomal dominant hyper-IgE syndrome. *Nature* 452, 773–776
- 193 Cai, T., Mazzoli, S., Meacci, F., Tinacci, G., Nesi, G., Zini, E. and Bartoletti, R. (2007) Interleukin-6/10 ratio as a prognostic marker of recurrence in patients with intermediate risk urothelial bladder carcinoma. *J. Urol.* 178, 1906–1911
- 194 Ishikawa, H., Tsuyama, N., Obata, M. and Kawano, M. M. (2006) Mitogenic signals initiated via interleukin-6 receptor complexes in cooperation with other transmembrane molecules in myelomas. *J. Clin. Exp. Hematop.* 46, 55–66
- 195 Knupfer, H. and Preiss, R. (2010) Serum interleukin-6 levels in colorectal cancer patients: a summary of published results. *Int. J. Colorectal Dis.* 25, 135–140
- 196 Vidal-Vanaclocha, F., Mendoza, L., Telleria, N., Salado, C., Valcarcel, M., Gallot, N., Carrascal, T., Egilegor, E., Beaskoetxea, J. and Dinarello, C. A. (2006) Clinical and experimental approaches to the pathophysiology of interleukin-18 in cancer progression. *Cancer Metastasis Rev.* 25, 417–434
- 197 Melisi, D., Niu, J., Chang, Z., Xia, Q., Peng, B., Ishiyama, S., Evans, D. B. and Chiao, P. J. (2009) Secreted interleukin-1 $\alpha$  induces a metastatic phenotype in pancreatic cancer by sustaining a constitutive activation of nuclear factor- $\kappa$ B. *Mol. Cancer Res.* 7, 624–633
- 198 McCarron, S. L., Edwards, S., Evans, P. R., Gibbs, R., Dearnaley, D. P., Dowe, A., Southgate, C., Easton, D. F., Eeles, R. A. and Howell, W. M. (2002) Influence of cytokine gene polymorphisms on the development of prostate cancer. *Cancer Res.* 62, 3369–3372
- 199 Singh, R. K. and Varney, M. L. (2000) IL-8 expression in malignant melanoma: implications in growth and metastasis. *Histol. Histopathol.* 15, 843–849
- 200 Brat, D. J., Bellail, A. C. and Van Meir, E. G. (2005) The role of interleukin-8 and its receptors in gliomagenesis and tumoral angiogenesis. *Neuro Oncol.* 7, 122–133
- 201 Artl, A., Vorndamm, J., Muerkoster, S., Yu, H., Schmidt, W. E., Folsch, U. R. and Schafer, H. (2002) Autocrine production of interleukin 1 $\beta$  confers constitutive nuclear factor  $\kappa$ B activity and chemoresistance in pancreatic carcinoma cell lines. *Cancer Res.* 62, 910–916
- 202 Xu, S. and Lam, K. P. (2001) B-cell maturation protein, which binds the tumor necrosis factor family members BAFF and APRIL, is dispensable for humoral immune responses. *Mol. Cell Biol.* 21, 4067–4074
- 203 Sparmann, A. and Bar-Sagi, D. (2004) Ras-induced interleukin-8 expression plays a critical role in tumor growth and angiogenesis. *Cancer Cell* 6, 447–458
- 204 Saijo, Y., Tanaka, M., Miki, M., Usui, K., Suzuki, T., Maemondo, M., Hong, X., Tazawa, R., Kikuchi, T., Matsushima, K. and Nukiwa, T. (2002) Proinflammatory cytokine IL-1  $\beta$  promotes tumor growth of Lewis lung carcinoma by induction of angiogenic factors: *in vivo* analysis of tumor-stromal interaction. *J. Immunol.* 169, 469–475
- 205 Kimsey, T. F., Campbell, A. S., Albo, D., Wilson, M. and Wang, T. N. (2004) Co-localization of macrophage inflammatory protein-3 $\alpha$  (Mip-3 $\alpha$ ) and its receptor, CCR6, promotes pancreatic cancer cell invasion. *Cancer J.* 10, 374–380
- 206 Porcile, C., Bajetto, A., Barbero, S., Pirani, P. and Schettini, G. (2004) CXCR4 activation induces epidermal growth factor receptor transactivation in an ovarian cancer cell line. *Ann. N.Y. Acad. Sci.* 1030, 162–169
- 207 Kundu, N. and Fulton, A. M. (2002) Selective cyclooxygenase (COX)-1 or COX-2 inhibitors control metastatic disease in a murine model of breast cancer. *Cancer Res.* 62, 2343–2346
- 208 Chang, S. H., Ai, Y., Breyer, R. M., Lane, T. F. and Hla, T. (2005) The prostaglandin E2 receptor EP2 is required for cyclooxygenase 2-mediated mammary hyperplasia. *Cancer Res.* 65, 4496–4499

- 209 Subbarayan, V., Sabichi, A. L., Llansa, N., Lippman, S. M. and Menter, D. G. (2001) Differential expression of cyclooxygenase-2 and its regulation by tumor necrosis factor- $\alpha$  in normal and malignant prostate cells. *Cancer Res.* 61, 2720–2726
- 210 Denkert, C., Kobel, M., Berger, S., Siegert, A., Leclere, A., Trefzer, U. and Hauptmann, S. (2001) Expression of cyclooxygenase 2 in human malignant melanoma. *Cancer Res.* 61, 303–308
- 211 Goulet, A. C., Einsphar, J. G., Alberts, D. S., Beas, A., Burk, C., Bhattacharyya, A., Bangert, J., Harmon, J. M., Fujiwara, H., Koki, A. and Nelson, M. A. (2003) Analysis of cyclooxygenase 2 (COX-2) expression during malignant melanoma progression. *Cancer Biol. Ther.* 2, 713–718
- 212 Souza, R. F., Shewmake, K., Beer, D. G., Cryer, B. and Spechler, S. J. (2000) Selective inhibition of cyclooxygenase-2 suppresses growth and induces apoptosis in human esophageal adenocarcinoma cells. *Cancer Res.* 60, 5767–5772
- 213 Wild, P. J., Kunz-Schughart, L. A., Stoehr, R., Burger, M., Blaszyk, H., Simon, R., Gasser, T., Mihatsch, M., Sauter, G. and Hartmann, A. (2005) High-throughput tissue microarray analysis of COX2 expression in urinary bladder cancer. *Int. J. Oncol.* 27, 385–391
- 214 Farrow, B. and Evers, B. M. (2002) Inflammation and the development of pancreatic cancer. *Surg. Oncol.* 10, 153–169
- 215 Mendes, R. A., Carvalho, J. F. and Waal, I. (2009) An overview on the expression of cyclooxygenase-2 in tumors of the head and neck. *Oral Oncol.* 45, e124–e128
- 216 Su, J. L., Shih, J. Y., Yen, M. L., Jeng, Y. M., Chang, C. C., Hsieh, C. Y., Wei, L. H., Yang, P. C. and Kuo, M. L. (2004) Cyclooxygenase-2 induces EP1- and HER-2/Neu-dependent vascular endothelial growth factor-C up-regulation: a novel mechanism of lymphangiogenesis in lung adenocarcinoma. *Cancer Res.* 64, 554–564
- 217 Heuze-Vourc'h, N., Zhu, L., Krysan, K., Batra, R. K., Sharma, S. and Dubinett, S. M. (2003) Abnormal interleukin 10R $\alpha$  expression contributes to the maintenance of elevated cyclooxygenase-2 in non-small cell lung cancer cells. *Cancer Res.* 63, 766–770
- 218 Forones, N. M., Kawamura, K. Y., Segreto, H. R., Artigiani Neto, R., Focchi, G. R. and Oshima, C. T. (2008) Expression of COX-2 in stomach carcinogenesis. *J. Gastrointest. Cancer* 39, 4–10
- 219 Tang, X., Sun, Y. J., Half, E., Kuo, M. T. and Sinicrope, F. (2002) Cyclooxygenase-2 overexpression inhibits death receptor 5 expression and confers resistance to tumor necrosis factor-related apoptosis-inducing ligand-induced apoptosis in human colon cancer cells. *Cancer Res.* 62, 4903–4908
- 220 Zhang, L., Zhang, W. P., Hu, H., Wang, M. L., Sheng, W. W., Yao, H. T., Ding, W., Chen, Z. and Wei, E. Q. (2006) Expression patterns of 5-lipoxygenase in human brain with traumatic injury and astrocytoma. *Neuropathology* 26, 99–106
- 221 Ye, Y. N., Wu, W. K., Shin, V. Y., Bruce, I. C., Wong, B. C. and Cho, C. H. (2005) Dual inhibition of 5-LOX and COX-2 suppresses colon cancer formation promoted by cigarette smoke. *Carcinogenesis* 26, 827–834

---

**Received 19 November 2010/1 March 2011; accepted 7 March 2011**

**Published on the Internet 26 September 2011, doi 10.1042/BSR20100136**

---